

Review

Hydrogels for Cartilage Regeneration, from Polysaccharides to Hybrids

Daniela Anahí Sánchez-Téllez ^{1,2,*} , Lucía Téllez-Jurado ¹ and
Luis María Rodríguez-Lorenzo ^{2,3} 

¹ Instituto Politécnico Nacional-ESIQIE, Depto. Ing. en Metalurgia y Materiales, UPALM-Zacatenco, Mexico City 07738, Mexico; ltellezj@ipn.mx

² Networking Biomedical Research Centre in Bioengineering, Biomaterials and Nanomedicine, Centro de Investigación Biomédica en Red—Bioingeniería, Biomateriales y Nanomedicina (CIBER-BBN), Av. Monforte de Lemos 3-5, Pabellón 11, Planta 0, 28029 Madrid, Spain; luis.rodriguez-lorenzo@ictp.csic.es

³ Department Polymeric Nanomaterials and Biomaterials, ICTP-CSIC, Juan de la Cierva 3, 28006 Madrid, Spain

* Correspondence: danielatellez06@gmail.com; Tel.: +34-695-198-353 or +521-55-29-54-32-91

Received: 14 October 2017; Accepted: 29 November 2017; Published: 4 December 2017

Abstract: The aims of this paper are: (1) to review the current state of the art in the field of cartilage substitution and regeneration; (2) to examine the patented biomaterials being used in preclinical and clinical stages; (3) to explore the potential of polymeric hydrogels for these applications and the reasons that hinder their clinical success. The studies about hydrogels used as potential biomaterials selected for this review are divided into the two major trends in tissue engineering: (1) the use of cell-free biomaterials; and (2) the use of cell seeded biomaterials. Preparation techniques and resulting hydrogel properties are also reviewed. More recent proposals, based on the combination of different polymers and the hybridization process to improve the properties of these materials, are also reviewed. The combination of elements such as scaffolds (cellular solids), matrices (hydrogel-based), growth factors and mechanical stimuli is needed to optimize properties of the required materials in order to facilitate tissue formation, cartilage regeneration and final clinical application. Polymer combinations and hybrids are the most promising materials for this application. Hybrid scaffolds may maximize cell growth and local tissue integration by forming cartilage-like tissue with biomimetic features.

Keywords: cartilage regeneration; polymeric hydrogels; polysaccharides; hybrid hydrogels; hybrid scaffolds

1. Introduction: Current Clinical Approaches and the Need for New Developments

The aim of this paper is to review the current state of the art of materials for cartilage substitution and regeneration. Section 1 describes the current state of the art in clinical treatments. Polymeric hydrogels used in cartilage regeneration and the reasons hindering their clinical success are reviewed in Section 2. The preparation techniques using polysaccharides and the resulting hydrogel properties are described in Section 3. Finally, future trends are explored in Section 4.

Several reviews about hydrogels for cartilage regeneration have been published for the last 10 years, focusing on preparation and characterization, natural and synthetic polymer precursors, gelation kinetics, cell and drug delivery, growth factors, mechanical properties and biocompatibility. Nevertheless, most of those reviews do not propose new alternatives to improve hydrogels properties which can fulfill the real clinical needs in terms of tissue regeneration, mechanical properties and degradation kinetics. Therefore, this paper reviews relevant literature published during 2013–2017, related to the application, fabrication, characterization, in vitro and in vivo assays of biomaterials

based on hydrogels for cartilage regeneration. The studies selected are articles and reviews written in English.

Currently, there are no clinical satisfactory solutions for cartilage tissue regeneration [1–3]. The most widely used clinical procedure to heal cartilage injury involves penetrating the wound to the subchondral bone, allowing the access of blood flow and new biological material [4–7]. However, such clinical treatments often result in the formation of fibrocartilaginous tissue (Figure 1), which is weaker than the original one, failing to integrate properly with surrounding tissue, and degrading over a period between 6 to 12 months [8–12].

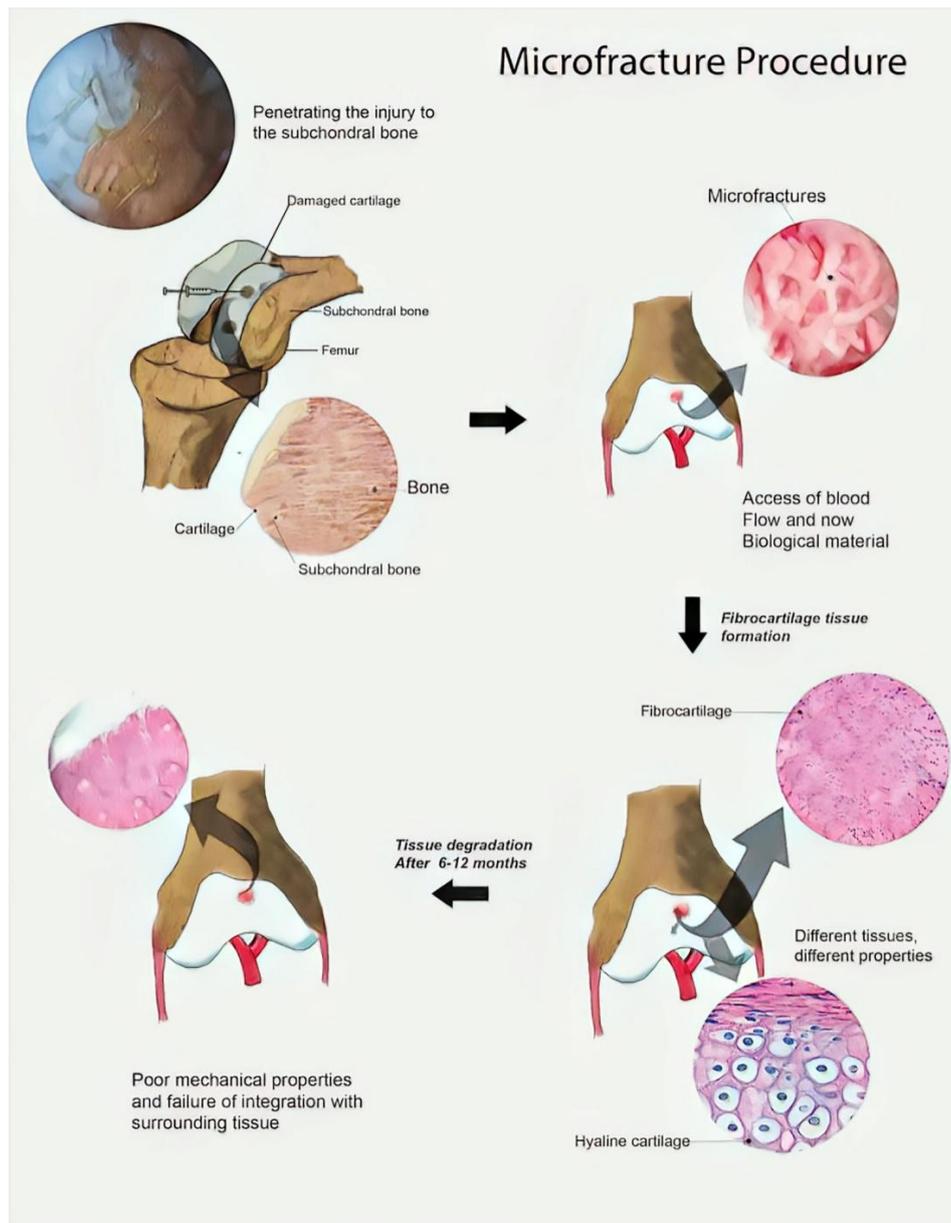


Figure 1. Common clinical procedure used to heal cartilage injury. Original illustration designed and provided by the authors.

During the last few years, material scientists and tissue engineers have tried to help clinicians by confronting the challenge of manufacturing porous 3D scaffolds which resemble the chemical composition and architecture (Figure 2) of the extracellular matrix (ECM) of the cartilage [13–19]. Most

of the studies are directed to determine how chemical composition and architecture influence cellular phenotype, differentiation, integration and extracellular matrix secretion during in vitro [20–25] and in vivo [26–32] assays.

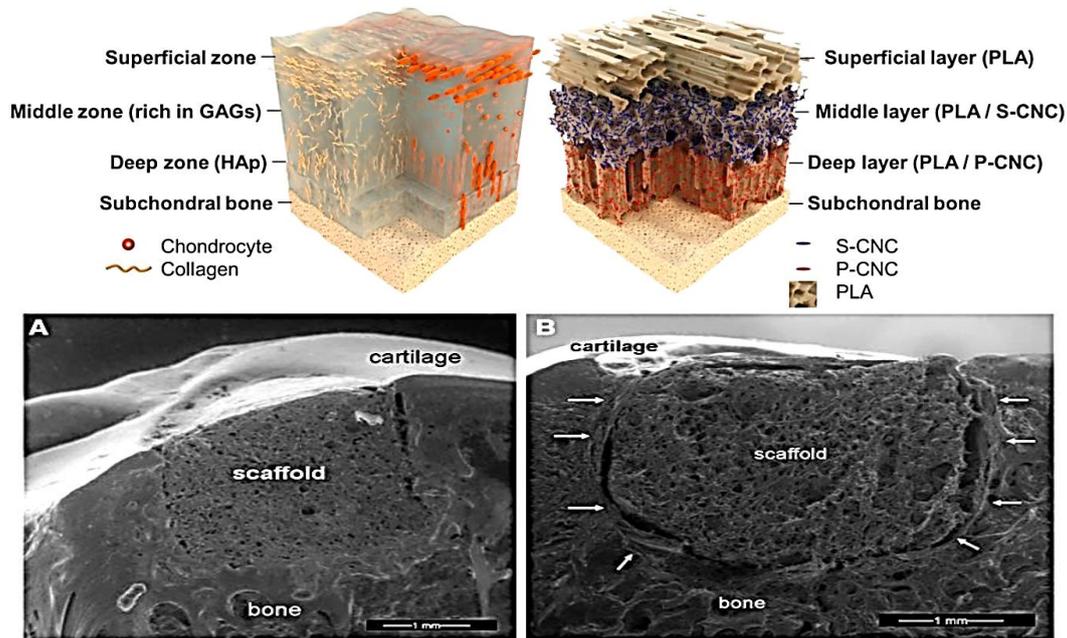


Figure 2. Top. How artificial scaffolds try to mimic the anisotropic characteristic in cartilage tissue [33,34]. Reprinted with permission of [33] Sandra Camarero-Espinosa et al. *Biomaterials*, 74: 42–52. Bottom. Matrix regeneration within a macroporous non-degradable implant for osteochondral defects is not enhanced with partial enzymatic digestion of the surrounding tissue. (A) Environmental scanning electron microscopy of a longitudinal slice taken through the cartilage-bone-implant. The specimen was retrieved at the 1 month post-operative time point. Good integration between the implant and the surrounding bone and articular cartilage was observed. (B) Environmental scanning electron microscopy image of a longitudinal slice taken through the cartilage-bone-implant construct at 3 months. Fibrous encapsulation of the implant is highlighted by arrows. Reprinted with permission of [34] Aaron J. Krych et al. *J. Mater. Sci. Mater. Med.*, 24: 2429–2437.

Several natural and synthetic polymers are being used to create novel materials, as collected in Table 1, in attempts to produce scaffolds for tissue engineering and regenerative medicine having clinical application [35].

Table 1. Scaffolds in clinical and preclinical use for cartilage regeneration.

Application	Material	Problem-result	Ref.
Nose (dorsal augmentation material in rhinoplasty)	Tissue-engineered chondrocyte PCS (Porcine Cartilage-derived Substance) scaffold construct.	Preliminary animal study: Excellent biocompatibility, neocartilage formation starts. However, it was not confirmed that the constructs contributed to the formation of neocartilage.	[36]

Table 1. Cont.

Application	Material	Problem-result	Ref.
Knee (subchondral bone)	Osteochondral biomimetic nanostructured scaffold Maioregen [®]	Better results in healing complex lesions in comparison with the implantation of a purely chondral scaffold.	[37]
	Cell-free biphasic scaffold: collagen-hydroxyapatite osteochondral scaffold	Statistically significant improvement in clinical scores. At 5 years, between 60.9% and 78.3% of the cases showed complete filling of the cartilage, complete integration of the graft, intact repaired tissue surface and homogeneous repaired tissue.	[38]
	Nanostructured biomimetic three-phasic collagen-hydroxiapatite construct	The implantations to treat chondral and osteochondral knee defects were effective in terms of clinical outcome, although MRI detected abnormal findings.	[39]
Knee (chondral defects)	Autologous ovine MNC Cell-seeded and cell-free DL-poly-lactide-co-glycolide (PLGA) scaffolds	The engineered tissue had not local or systemic adverse effects. However, only a poor integration of the tissue engineering product into adjacent tissue was reached and the formed ECM was not mature enough for long-lasting weight-loading resistance.	[40]
	Type I collagen-hydroxyapatite (Maioregen [®]) nanostructural biomimetic osteochondral scaffold	The use of the Maioregen [®] scaffold is a good procedure for the treatment of large osteochondral defects; however, the lesion site seems to influence the results. Patient affected in the medial femoral condyle showed better results.	[41]
	DeNovo (RevaFlex) engineered tissue graft	Preliminary evidence suggests that DeNovo ET implant is capable of spontaneous matrix formation with no immune response, improving function and recreating hyaline-like cartilage.	[42]
Knee (femoral condyles)	Biphasic cylindrical osteochondral composite construct of DL-poly-lactide-co-glycolide (PLGA). Its lower body is impregnated with β -tricalcium phosphate (TCP)	The regenerated osteochondral tissue was evaluated as a tissue of acceptable quality. Regenerated cartilage was defined as being hyaline when the ground substance was homogeneous without fibrous texture.	[43]
Tibial plateau (osteochondral scaffold)	Osteochondral biomimetic collagen-hydroxiapatite scaffold (Maioregen [®] , Fin-ceramica, Faenza, Italy)	MRI abnormalities. Clinical outcome with stable results up to a mid-term follow-up.	[44]

Table 1. Cont.

Application	Material	Problem-result	Ref.
Microfractured defect (for filling microfractures)	BioCartilage™, product containing dehydrated, micronized allogeneic cartilage, implanted with the addition of platelet rich plasma	No human clinical outcomes data available. Data regarding results are limited to expert opinion.	[45]
	Chondroitin sulfate adhesive-Poly(ethylene glycol) diacrylate (PEGDA) hydrogel system combined with standard microfracture surgery	Significant increase in tissue fillers with defects in a short-term follow-up.	[46]
Knee (for donor site filling)	Artificial TruFit cylinders made of fully synthetic material called PolyGraft®-Material: 50% copolymer (PDLG), composed of 85% poly(D,L-lactide) and 15% glycolide; 40% calcium sulphate, 10% PGA fibers	No clinical improvement could be found. The regeneration of the filled defects took more than 2 years, even though TruFit Plugs are supposed to stimulate cartilage and bone cell migration from the surrounding tissue to the synthetic cylinders.	[47]
	Porous poly(ethylene oxide)terephthalate/butylene terephthalate (PEOT/PBT) implants	Treated defects did not cause postoperative bleeding. Well integration. Surface stiffness was minimally improved compared to controls. Considerable biodegradation after 9 months. Congruent fibrocartilaginous surface repair with interspersed fibrous tissue formation in implanted sites. Donor site: fibrocartilaginous surface repair.	[48]
Shoulder	Engineered hyaluronic acid membrane, Hyalograft®	Using the hyaluronic membrane had no effect on the final outcome. No difference was observed between the fibrocartilage tissue formed after implementing microfractures and the fibrocartilage tissue grown on the hyaluronic acid membrane scaffold.	[49]

However, the complexity of the physical structure and properties of cartilage, including mechanical [50–54], anisotropic [55–57], nonlinear [58–60], inhomogeneous [61–63] and viscoelastic properties [64–66], are thought to be directly related to the failing of most of the attempts made to fabricate artificial substitutes for cartilage [67–71]. As a consequence, there are not yet biomaterials for cartilage regeneration in clinical use with satisfactory results. Scaffolds based on tissue-engineered constructs, osteochondral biomimetic scaffolds, cell-free biphasic or three-phasic scaffolds, autologous scaffolds, engineered-tissue grafts, porous implants have not demonstrated to be a satisfactory solution in clinical application. Therefore, it seems evident that there is a need to design more suitable scaffolds and to develop new types of materials which can be used for cartilage regeneration.

In the case of articular cartilage repair, the required materials must provide successful mechanical properties, biological delivery, fixation of the device in situ and stability to the joint. Besides, the assays done to these materials need to be based on the intended biological effect and potential risks

which have to be evaluated, such as toxicity, dedifferentiation, immunogenicity and contamination. Some preclinical trials with little animal models, such as rats and white New Zealand rabbits, are necessary to predict how biomaterials may behave during clinical assays. Moreover, some preliminary studies can be done in cadaveric human bodies; however extensive trials with large animal preclinical models are mandatory to obtain market approval. Therefore, just a few of the novel materials or tissue protocols are allowed to have clinical application or even to be commercialized. When trying to compare different studies of novel materials once they are introduced into clinical practice, there are other problems: (1) lack of homogeneity due to the different studied population; (2) short- and mid-follow-ups; (3) use of different evaluation systems; (4) new scaffold-based strategies for cartilage regeneration, either cell seeded or cell-free biomaterials; (5) procedures which differ in scaffold fixation methods, surgical approaches and postoperative rehabilitation phases. Therefore, there is the need among scientists, clinicians, industry and regulatory experts to improve communication and collaboration in order to overcome all the barriers in tissue engineering and to establish a defined road map to reach clinical application.

2. Hydrogels in Cartilage Regeneration

Hydrogels are emergent candidates for applications in cartilage regeneration. Hydrogels are three-dimensional hydrophilic polymer networks made up of water-soluble polymers, crosslinked by either covalent or physical methods [72] (Figure 3) to form a water-insoluble hydrogel [73,74]. Hydrogels can be composed of natural polysaccharides [75–77], proteins [78–82] or synthetic polymers [83–85]. Hydrogels are able to swell and retain great portions of water, from 20% to 99% by weight, when placed in aqueous solutions [86–88]. Hydrogels, tested as matrices to build up scaffolds, provide highly desirable 3D environments for cell growth, holding a great promise for the regeneration of cartilaginous tissue as *in vitro* and *in vivo* studies showed [89–94]. Several studies use cells to catalyze tissue formation while being distributed in 3D hydrogel matrices. Cell matrix adhesion to hydrogel is an important interaction which regulates stem cell survival, self-renewal, and differentiation. Using 3D culture systems (hydrogels) may provide an appropriate niche, scaffolding and environmental bioactive signals for cells. Depending on their physical structure and chemical composition, hydrogels can preserve a compositional and mechanical similarity with the native extracellular matrix of cartilage [95–98]. These properties are necessary for controlling cell response, differentiation and functional tissue regeneration [99–106].

One important reason for the choice of hydrogels in cartilage applications is the possibility of making them injectable which offers advantages over solid scaffolds such as the possibility of using a non-invasive approach. Injectable hydrogels can fill any shape defect and they may provide a homogeneous cell distribution within any defect size or shape prior to gelation [107–111]. Over recent years, a variety of naturally [112–119] and synthetically [120–122] derived materials such as silk [123,124], resilin [125], chitosan [126], chondroitin sulfate [127], hyaluronic acid [128–131], gelatin [132], agarose [133], alginate [134] poly(vinyl alcohol) (PVA) [95,135] poly(acrylic acid) [136], acrylamide [137] and many others have been used to form injectable hydrogels for cartilage repair.

Collagen II and glycosaminoglycans (GAGs) are cartilage-specific extracellular matrix components; they play a crucial role in regulating the expression of chondrocytic phenotype and in supporting chondrogenesis. They have been used for *in vitro* and *in vivo* assays [138]; many attempts have been made with different GAGs precursors of hydrogels to provide an appropriate biochemical and biomechanical environment for cells [139–146]. Unfortunately, hydrogels derived from GAGs degrade really fast, thus different chemical modifications have to be introduced. Several crosslinking degrees are necessary in attempt to modulate their degradation kinetics. However, their biological response is also modified [147–151]. In general, the studies found on literature can be divided into two main types: (1) cell-free hydrogel scaffolds; and (2) cell-seeded hydrogel scaffolds.

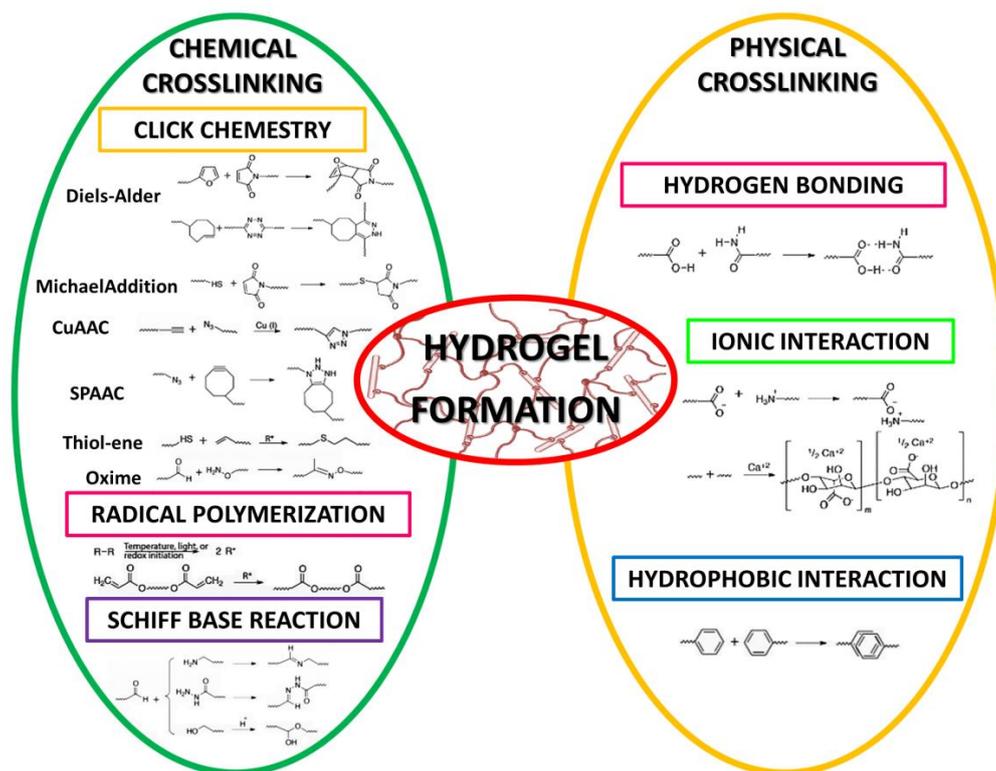


Figure 3. Chemical and physical crosslinking methods used to form hydrogels.

2.1. Cell-Free Hydrogel Scaffolds

Investigations based on cell-free hydrogel scaffolds focus on their physico-chemical characterization and mechanical properties [152]. These studies allow a full understanding of physical and chemical interactions within materials and how these interactions may affect biological and mechanical properties of scaffolds.

Poly(ethylene glycol) (PEG) and derived hydrogels are ones of the most widely used synthetic polymer for tissue engineering. The modulus for bovine articular cartilage, measured in compression mode, is 950 KPa, which is close to the value of the fully hydrated polyethylene glycol diacrylate (PEG-DA) hydrogel [87]. Polyvinyl alcohol is another synthetic polymer widely used to form hydrogels due to their excellent biocompatibility, high permeability to fluids (showing an equilibrium water content of $32 \pm 5\%$) and low friction coefficients (μ) in the range of 0.02 to 0.05 against smooth and wet substances. Some studies on PVA-based scaffolds aim to demonstrate that under tribological loading, friction and wear characteristics compatible to natural articular cartilage can be achieved [153]. As low friction coefficients are required for engineered cartilage, polyvinyl alcohol (PVA)/polyvinylpyrrolidone (PVP) hydrogels were synthesized with different polymerization degrees: 1700, 2400 and 2600 for the PVA; and different polymer concentrations: 10% *w/w*, 15% *w/w* and 20% *w/w* of PVA/PVP. It was found that the inner structures of hydrogels tend to be denser when polymer concentration and polymerization degree of PVA increase. While the friction coefficient increases (from 0.037 to 0.044) with an increment in the polymerization degree of PVA (average increase rate is approximate 3%), the friction coefficient decreases (from 0.033 to 0.03 for a 2.5 N load; from 0.049 to 0.045 for a 7.5 N load) with an increment in the polymer concentration (from 10% to 20%) in the low load region and under liquid lubrication. Thus, there is the need to keep friction coefficients stable under lubricated conditions [154].

Another study using PVA-based hydrogels, crosslinked with trimetaphosphate (STMP), revealed that fully hydrated hydrogels were covalently crosslinked systems when mechanically tested, with a rheological behavior (the G' changed from 0.01 MPa (0.01 Hz) to 0.02 MPa (15 Hz)) similar to that of

tibia cartilage ($G' = 0.03$ for tissue surface and $G' = 0.11$ for overall tissue) [86]. As previously said, it is an important challenge to develop scaffolds which possess mechanical properties mimicking those of cartilage tissue, since cartilage is a complex nonlinear, viscoelastic and anisotropic extracellular matrix structure. T. Chen et al. [155] reported that hydrodynamic conditions, simulating the motion-induced flow fields between the articular surfaces within the synovial joint, induce the formation of a distinct superficial layer on tissue engineered cartilage hydrogels. These hydrodynamic conditions enhance, on the superficial layers, the production of cartilage matrix proteoglycan, type II collagen and a highly aligned fibrillary matrix which resembles the alignment pattern in native tissue surface zone.

Since many materials do not exhibit a low friction coefficient or withstand several loading cycles, some of them are infiltrated with an interpenetrating network hydrogel to form functional scaffolds which provide load-bearing and tribological properties, similar to native cartilage ones. For example: (1) a porous three-dimensionally woven poly(ϵ -caprolactone) fiber scaffold was infused with a “tough-gel” made of alginate and polyacrylamide [83]; (2) a boundary lubricant functionalized PVA-based hydrogel was developed to be used as a synthetic replacement for focal defects in articular cartilage [156]. Other techniques to develop hydrogels with high mechanical strength are: by using a double network or two-step polymerization [88,157]; or by functionalizing hydrogels with different organic or inorganic molecules [158].

2.2. Cell-Seeded Hydrogel Scaffolds

Cell-based hydrogel scaffold therapy is one of the main strategies being investigated in cartilage regeneration. Several scaffolds and materials are being evaluated. These studies focus on whether or not the hydrogels provide an appropriate biochemical and biomechanical environment for a long-lasting hyaline-type cartilage regeneration. Decellularized extracellular matrices from natural tissues like dermis or adipose are being studied as functional biologic scaffolds (Figure 4). It is possible to ensure the bioactivity of a substrate when scaffolds are seeded with a specific cell type, either chondrocytes or stem cells, and if these cells are able to proliferate regardless of “natural” conditions. As an example, Giavaresi G. et al. [159] evaluated in vitro the biological influence of a decellularized human dermal extracellular matrix on human chondrocytes (NHAC-kn) and mesenchymal stromal cells (hMSC). The study showed that at 24 h after seeding, cells adhered consistently to dermal membranes (NHAC-kn = 93% and hMSC = 98%); at 7 days, cell viability index was 98% for both cell cultures seeded on dermal membranes; and after 14 days of culture, the indexes increased significantly for both cell cultures ($p < 0.0005$; NHAC-kn = 136% and hMSC = 263%). Furthermore, a biohybrid composite scaffold, composed by combining a decellularized Wharton’s jelly extracellular matrix with the polyvinyl alcohol (PVA)-based hydrogel, demonstrated its ability in promoting chondrocyte adhesion and scaffold colonization [160]. Other studies are examining extracellular matrices developed from porcine articular cartilage [161]. Although these substrates worked as proper scaffolds for the growth of cells, their therapeutic and functional efficiency in cartilage regeneration still need to be proved. Other cell therapies include implanting chondrogenic lines differentiated from mesenchymal stem cells (MSC) into different polysaccharides or synthetic hydrogels. Collagen hydrogels have proved to provide an appropriate 3D environment for MSC chondrogenesis, isolated from Wharton’s jelly of human umbilical cord, and to be cytocompatible matrices with great potential for cartilage engineering [162].

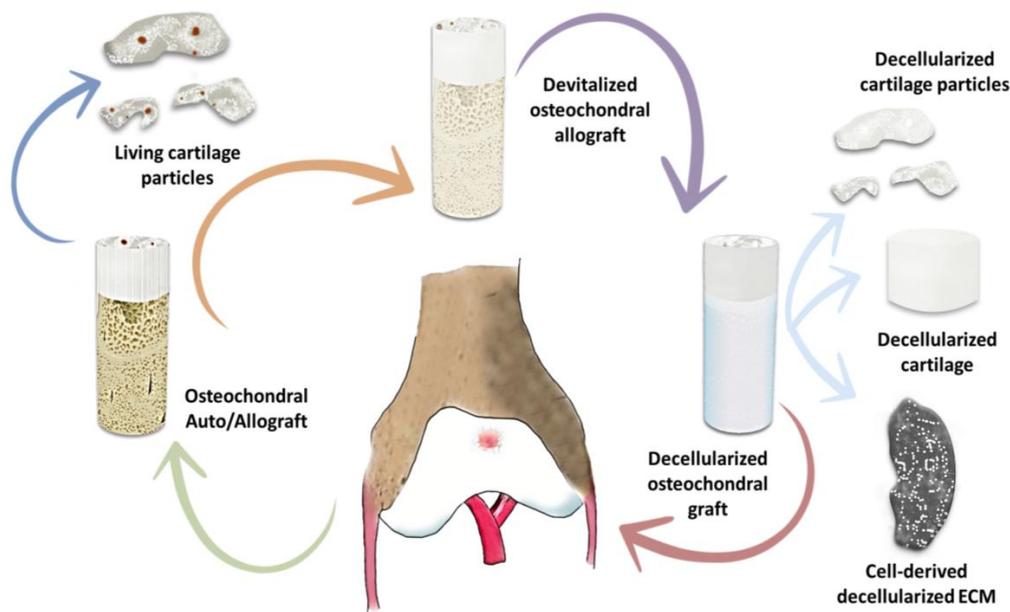


Figure 4. Matrix-based scaffold approaches for cartilage regeneration. Original illustration designed and provided by the authors.

One of the problems being reported when using cell therapies is the dedifferentiation of chondrocytes when cultured in two-dimensional cultures, making them less functional for cartilage repair. Wu L. et al. [163] hypothesized that functional exclusion of dedifferentiated chondrocytes can be achieved by detecting domains formation of collagen molecules deposited by chondrogenic cells into 3D environments. They reported a method which allows separation of functionally active chondrogenic cells, which produce high levels of collagen II, from functionally inferior dedifferentiated cells, which produce collagen X. To avoid dedifferentiation of cells once they are forming constructs, Lam J. et al. [164] investigated the ability of cell-laden bilayer hydrogels, by encapsulating chondrogenically and osteogenically pre-differentiated mesenchymal stem cells, by varying the period of chondrogenic pre-differentiation prior to implantation. Therefore, cell phenotype could be optimized in order to achieve ideal tissue repair. Furthermore, since regeneration of human articular cartilage is limited, various cellular sources have been studied, including adult and juvenile chondrocytes. Some studies have compared the formation of cartilage tissue, produced by juvenile, adult and osteoarthritic chondrocytes, inside 3D biomimetic hydrogels composed of poly(ethylene glycol) and chondroitin sulfate. It was found that after the cultured time, juvenile chondrocytes showed a greater upregulation of chondrogenic gene expression than adult chondrocytes, while OA chondrocytes showed a downregulation [101]. Another strategy being studied is the evaluation of therapeutic effects of intra-articular injections of hydrogels containing drugs used to treat osteoarthritis symptoms [165].

Some other studies are meant to analyze how the structure and fabrication methods of the scaffolds influence cells behavior. Due to their intimate contact with chondrocytes, scaffolds are important components of cell niche. Investigations into micro-architecture of scaffolds have revealed that mean pore size is cell-type specific and influences cellular shape, differentiation and extracellular matrix secretion (Figure 5). Studies in collagen-hyaluronic acid scaffolds, fabricated with different mean pore size, showed that scaffolds with the largest mean pore size (300 μm) stimulated significantly a higher cell proliferation, chondrogenic gene expression and cartilage-like matrix deposition [166]. When using synthetic materials to produce hydrogels, it is necessary to determine their in vitro pore size and mechanical stiffness after being rehydrated, in order to predict their in vivo behavior. Hui J. H. et al. [167] found that freeze-dried oligo[poly(ethylene glycol)fumarate] (OPF) hydrogels with

a pore size ranged from 20 to 433 μm in diameter and a mechanical stiffness of 1 MPa when rehydrated, enhance the formation of hyaline-fibrocartilaginous mixed tissue. However, these hydrogels, implanted alone into cartilage defects, are insufficient to generate a homogeneously hyaline cartilage repair tissue. Kwon H. et al. [168] demonstrated that scaffolds, with different pore size and fabrication methods, influence the microenvironment of chondrocytes and their response to proinflammatory substances. Having high levels of proinflammatory cytokines can cause cartilage destruction and instability of the engineered cartilage tissue. These authors found that silk scaffolds with larger pore sizes support higher levels of cartilage matrix and leach more efficiently proinflammatory cytokines into the medium, influencing cartilage gene expression.

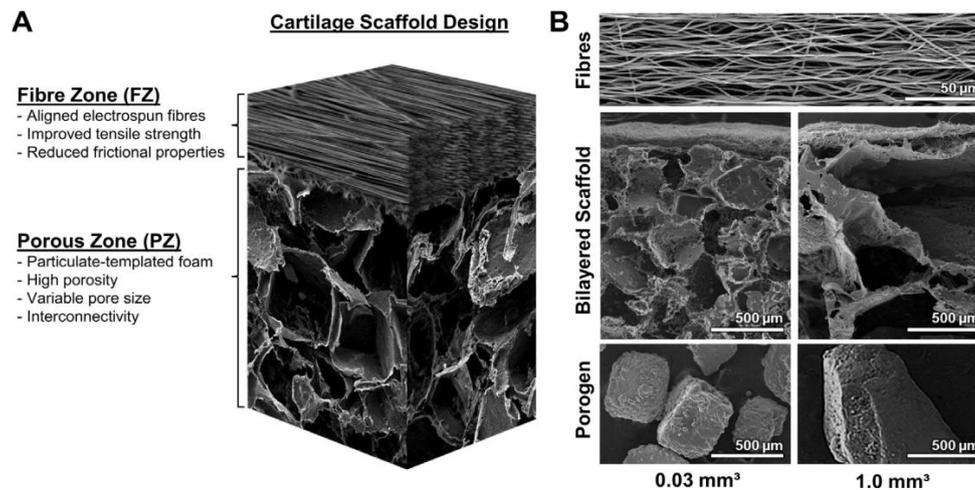


Figure 5. Bilayered cartilage scaffold (A) schematized by a diagram illustrating the electrospun fiber zone (FZ) deposited on a particulate-templated foam (PZ). The combination of the two distinct zones is designed to yield an anisotropic scaffold with a smooth articular surface and a more porous region for ECM deposition. (B) Electron microscopy images (top) of the aligned fiber zone that is shared between both scaffold varieties, (middle) the complete bilayered scaffolds with 0.3 mm^3 (left) and 1.0 mm^3 (right) pores, and (bottom) the sodium chloride porogens used to produce their respective scaffolds. Reprinted with permission from [169] J.A.M. Steele et al. Combinational scaffold morphologies for zonal articular cartilage engineering. *Acta Biomaterialia*. 10: 2065–2075.

Furthermore, each zone of cartilage tissue varies in regard to biochemical content, morphology and biomechanical function. Deeper cartilage zones present higher stiffness, higher proteoglycan concentration but lower cellular density and same collagen concentration along cartilage tissue. In a general structural perspective, cartilage can be simplified into two main regions: (1) the superficial zone which exhibits a high tensile strength and low friction coefficient to keep a smooth articulation; (2) a dense ECM region rich in proteoglycan molecules which give the tissue adequate compressive mechanical properties by producing a high osmotic pressure within the tissue. Therefore, when fabricating a bilayer or three-layer scaffold, pore size and fabrication method of each layer influence the microenvironment of chondrocytes. As shown by Steele J.A.M. et al. [169], tissue engineering scaffolds can be designed to vary in morphology and function, offering a template: (1) to mimic the structural organization and functional interface of cartilage superficial zone; (2) to increase the extracellular matrix production; (3) to enhance the anisotropic mechanical properties. These authors fabricated a multi-zone cartilage scaffold by electrostatic deposition of polymer microfibers onto particulate-templated scaffolds with 0.03 mm^3 and 1.0 mm^3 porogens. They demonstrated that bilayered scaffolds can closely mimic some of the structural characteristics of native cartilage due to: (1) the addition of aligned fiber membranes enhances the mechanical and surface properties of scaffolds; (2) zonal analysis of scaffolds showed region-specific variations in chondrocyte number, sulfated

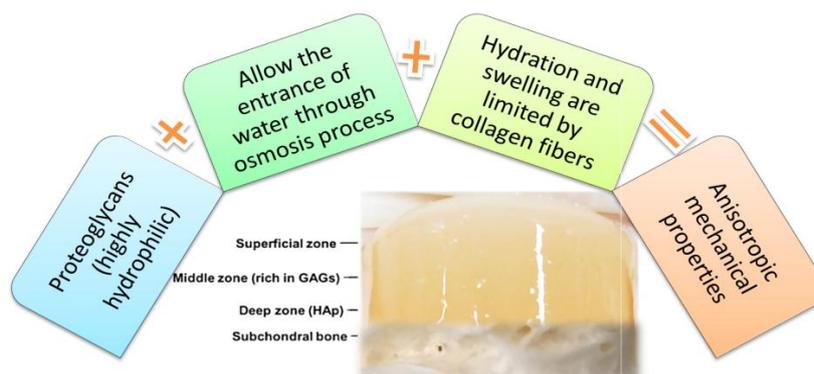
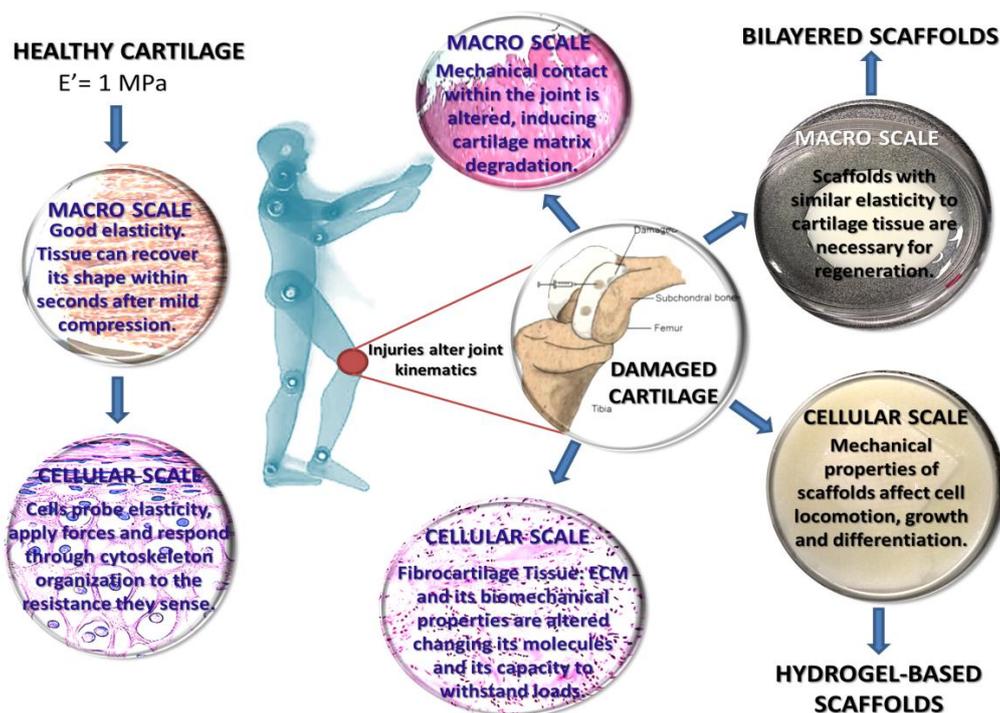
GAG-rich extracellular matrix and chondrocytic gene expression; (3) smaller porogens (0.03 mm^3) yield higher GAGs accumulation and aggrecan gene expression.

It is important to understand the multi-scale biomechanical behavior of cartilage tissue, in order to realize the connection among joint kinematics, tissue-level mechanics, cell mechanics and mechanotransduction, matrix mechanics and the nanoscale mechanics of matrix macromolecules. Therefore, understanding mechanical behavior at each scale helps to correlate cell biology, matrix biochemistry and tissue structure/function of cartilage (Figure 6).

Moreover, the combination of cell-based therapies with growth factor delivery, which can locally signal cells promoting their function, is also being investigated. Since morphogenetic protein (BMP-2) and transforming growth factor (TGF- β) are cytokines proposed as stimulants for cartilage repair, it is necessary to undertake a detailed comparative analysis of their biological effects on chondrocytes [170]. As an example, when chondrocytes are encapsulated in PEG hydrogels functionalized with transforming growth TGF- β 1, proliferation and matrix production increase, in comparison with cells in hydrogels where TGF- β 1 is dosed in the culture medium or untreated TGF- β 1 hydrogels [171]. In another study, chondroitin sulfate-bone marrow adhesive hydrogel was used to localize and carry BMP-2 protein, which enhance articular tissue formation. It was demonstrated that these hydrogels were capable of supporting articular chondrocytes, viability and phenotype retention, stimulating cells to produce hyaline-like extracellular matrix [172]. Although expensive growth factors in cultures are used, there is still a production of cartilage with inferior mechanical and structural properties compared with the natural tissue. However, recent evidence suggests that GAGs incorporated into tissue engineering scaffolds can isolate and/or activate growth factors, mimicking better the natural extracellular matrix [173]. For example, in the presence of TGF- β 3 releasing microspheres, gellan gum hydrogels facilitate a greater cell proliferation than fibrin or agarose hydrogels. Histological and biochemical analysis of these hydrogels indicated that fibrin hydrogel was the least chondro-inductive, while agarose and gellan gum hydrogels supported more robust cartilage formation because of a greater GAGs accumulation within the constructs. Unfortunately, gellan gum hydrogels stained more intensely for collagen type II and collagen type I, suggesting a fibrocartilaginous tissue phenotype [174]. There are other studies focused on calcified cartilage zone, which provides mechanical integration between articular cartilage and subchondral bone. Lee W.D. et al. [175] developed tissue-engineered osteochondral-like constructs with bone marrow stromal cells (BMSC), as single cell source. Cartilage tissue and a porous bone substitute substrate were formed with an interfacial zone of calcified cartilage. The authors found that the presence of calcified cartilage increased the shear load that the construct may withstand at the interface. However, preclinical studies are needed to determine if these osteochondral-like constructs could repair joint defects in vivo.

Evaluating cell-free and cell-based hydrogels reviewed above, only a few of these biomaterials have been used in clinical applications [45,46,74,176–179] because of four main unsolved problems in tissue engineering: (1) toxicity of some crosslinking agents [180]; (2) lack of mechanical integrity [156,181–187]; (3) poor control of gelation kinetics [188,189]; (4) unsuitable degradation kinetics [190–194].

Since some of the reactions used to synthesize hydrogels are limited due to their complexity, the use of cytotoxic reagents, instability of some functional groups, possible side reactions and low coupling efficiency, there is the need to explore and exploit simple and highly efficient methods which may be applicable to a great variety of biodegradable polymeric precursors.



Ref.	Scaffold Material	Mechanical Properties							
209	PCL control	Tensile Young's modulus E (MPa) = 2.09±0.21 Ultimate tensile strength (MPa) = 15.15±1.3 Compressive modulus (MPa) = 6.37±0.88							
	PCL scaffold + hyaluronic acid hydrogel	Tensile Young's modulus E (MPa) = 2.08±0.24 Ultimate tensile strength (MPa) = 14.25±0.99 Compressive modulus (MPa) = 6.54±0.75							
	Si-HPMC hydrogel	Compressive storage moduli E' (kPa) = 5±3							
214	Si-HPMC/Nanofibers (3% wt)	Compressive elastic modulus E' (kPa) = 27±2							
Ref.	Scaffold type	Porogens size (mm ³)	Tensile modulus (MPa)	Tensile stress at failure (MPa)	Compressive modulus (kPa)	Compressive stress (kPa) at 10% strain	Interfacial stiffness (aligned) (N/mm)	Interface peak load (aligned) (N)	Surface roughness (R _s ; μm)
169	Particulate-template PCL	0.03	0.04±0.02	0.05±0.01	40±5	5±1	N/A	N/A	16±10
		1.0	0.10±0.05	0.03±0.01	20±5	2±1	N/A	N/A	26±20
	Bilayered PCL	0.03	1.1±0.3	0.4±0.1	44±21	4.7±2.0	0.24±0.04	0.68±0.03	3.0±0.8
		1.0	1.6±0.5	0.4±0.2	14±4	2.0±0.4	0.23±0.002	1.15±0.03	2.1±0.5
	Fibre zone PCL	-	28±2	12±3	140±20	70±4	N/A	N/A	3±1

Figure 6. Correlation among the multi-scale biomechanical behavior of cartilage tissue. Original illustration designed and provided by the authors.

3. Polysaccharides Versus Synthetic Hydrogels

The degradation rate and mechanical properties of manufactured hydrogels must be compatible with the growth of new tissue [109]. As mentioned before, hydrogels can be made of either natural or synthetic polymers, each of them with advantages and disadvantages, or even a combination of both,

whether or not a reduction of disadvantages from the individual components can be obtained. When using natural precursors, good biocompatibility and bioactivity are ensured in the hydrogel scaffolds. However, there will be a high degradation rate, in contact with body fluids or medium, and a limited mechanical behavior, since natural polymer components are extracted from tissues and subsequently reconstructed to form hydrogels [173,195]. Nevertheless, the strength of these natural hydrogels can be increased by making the polymer matrix denser, using a chemical crosslinking or making chemical modifications. On the other hand, using synthetic precursors may provide appropriate physical and chemical properties for hydrogel scaffolds; however a good cell biological response and an adequate degradation rate may not occur [196,197]. The strength of the synthetic hydrogels can be raised by changing the molecular weight of the starting polymers, increasing polymer concentration or even the degree of functionalization, using reactive groups during the crosslinking reaction.

3.1. Manufacturing Techniques and Their Influence on Hydrogel Properties

Depending on the polymer precursors, hydrogels can be synthesized in different ways. In the first step a polymer is modified with a functional group; then the polymer is crosslinked, either physically or chemically, to form a three-dimensional structure. While chemical hydrogels are covalently crosslinked, physical hydrogels are not. Crosslinking can take place at the same time or after the copolymerization [198]. In situ crosslinked cytocompatible injectable hydrogels can be formed using: (1) non-toxic chemical crosslinkers, as in the Michael Addition Reaction, Click Chemistry, Schiff Base Reaction, and photo-crosslinking reactions; (2) enzymes for a biological crosslinking; (3) physical interactions, such as ionic and hydrophobic ones; (4) supramolecular chemistry utilizing self-assembly molecules [199].

The morphology and physico-chemical structure of hydrogels also depend on processing conditions applied during their formation, for example using electrospinning or cryogelation techniques. Hydrogel morphologies may range from fibrils, characteristic of protein-based hydrogels such as collagen and fibrin, to amorphous, characteristic of synthetic hydrogels such as PEG. When using the electrospinning technique, it is possible to obtain hydrogels with aligned fibrils morphologies. Mirahmadi F. et al. [200] added degummed chopped silk fibers and electrospun silk fibers to thermosensitive chitosan/glycerophosphate hydrogels, to reinforce scaffolds for hyaline cartilage regeneration. The results showed that mechanical properties of hydrogels were significantly enhanced; besides the composition of the scaffolds supported the chondrogenic phenotype. Nevertheless, when using cryogelation, because of ice crystals, a controlled porosity can be induced into hydrogels, helping them recovering their shape [201]. Another method to fabricate three-dimensional porous hybrid scaffolds for articular cartilage repair is combining freeze-dried [192,202,203] natural components with synthetic polymers, provide scaffolds with mechanical strength and an environment similar to natural ECM, to let chondrocytes proliferate. Lyophilization or freeze-drying technique produces highly porous structures with open pores throughout the scaffolds. The pores are introduced into the scaffolds, first by ice crystal formation, then by freeze-drying them. For this reason pore architecture reflects the ice crystal morphology [203]. Novel collagen/poly(lactide) (PLA), chitosan/PLA, and collagen/chitosan/PLA hybrid scaffolds were fabricated by freeze-drying technique [204]. It was observed that collagen binds water inside the scaffold structure and it helps cells to penetrate into the hybrid scaffolds. To enhance anisotropic properties of cartilage scaffolds, aligned unidirectional pores can be formed depending on the real alignment of cells and the type of extracellular matrix that has to be repaired. Collagen-hybrid scaffolds, constructed by directional freezing, were studied. When varying freezing rates and suspension media, it is possible to obtain collagen-hybrid scaffolds with unidirectional pores, tunable pore sizes and pore morphologies [203]. The results demonstrated that directed horizontal ice dendrite formation and vertical ice crystal nucleation are responsible for aligned unidirectional ice crystal growth and, consequently, for aligned unidirectional pore structure of the collagen-hybrid scaffolds.

Since conventional fabrication techniques may not provide a precisely control of pore size, interconnectivity or pore geometry for scaffolds, solid freeform fabrication (SFF) techniques are now being used to produce 3D scaffolds with an organized interconnected pore structure which ensures good functionality and good mechanical strength, necessary to maintain new cartilage formation.

Bioprinting and plotting are being used as freeform fabrication techniques. These emerging techniques are used to fabricate viable 3D tissue constructs through a precise deposition of cells in hydrogels [205]. However, scaffolds, fabricated by these printing systems, often lack of flexibility and adequate mechanical properties [138,206]. Xu T. et al. [207] described the construction of a hybrid inkjet printing/electrospinning system that can be used to fabricate viable cartilage engineered tissue. They fabricated a five-layer construct, 1 mm thick, made of electrospun polycaprolactone fibers alternated with inkjet printing of rabbit elastic chondrocytes suspended in a fibrin-collagen hydrogel. One week after printing, evidence showed more than 80% of cell viability, cell proliferation and formation of cartilage-like tissue in the five-layer construct, both in vitro and in vivo assays, and demonstrated an improvement of mechanical properties, in comparison with printed alginate or fibrin-collagen hydrogels.

Novel techniques of tissue scaffold fabrication, as ultrafast pulse DLW lithography, are attractive due to their 3D structuring capability, spatial resolution, scaling flexibility and diversity of working materials.

3.2. Degradation Kinetics, Physical Properties (Applicability) and Biological Effects

Hydrogels degradation kinetics should be compatible with new tissue formation kinetics, in order to ensure a good integration of the construct. If hydrogels degrade very fast, it will trigger occurrence of defects in the formed tissue, such as cysts. On the other hand, if a very slow degradation occurs, hydrogels will inhibit the formation of new biological material and their integration with the surrounding tissue. Degradation of hydrogels can take place by either hydrolytic or enzymatic mechanisms. The hydrolytic degradation happens when hydrogel is kept in contact with fluids by breaking the polymer chains or the crosslinked network. This type of degradation mechanism can be controlled by limiting the amount of degradable precursor, used to synthesize hydrogels. The enzymatic degradation is caused by cells when they begin to invade the hydrogel or when encapsulated cells within the hydrogel start to proliferate or migrate throughout it. Many natural origin proteins have sites of cleavage in the protease, allowing hydrogels to degrade during the replacement of the ECM. This type of degradation may also depend on the degree and the type of chemical crosslinking in the hydrogels used as scaffolds [201].

3.3. Specificities of Polysaccharide-Based Hydrogels

When trying to manufacture biomimetic scaffolds for cartilage tissue regeneration, naturally-derived hydrogels are widely used, due to their macromolecular properties and because the employed biopolymers are part of the natural tissue that needs to be healed. Most of the studied naturally-derived hydrogels are based on biopolymers such as collagen, gelatin, chitosan, hyaluronic acid, chondroitin sulfate, agarose, alginate and fibrin [109,208].

3.4. Biological Response of Polysaccharide-Based Hydrogels

Since hyaluronic acid is a fundamental component of natural cartilage matrix, some studies have shown its importance and its good qualities as an excellent naturally derived polymer. Injections of hyaluronan into osteoarthritic joints have proved to restore viscoelasticity, augment joint fluid flow, normalize endogenous hyaluronan synthesis, and provide joint function [109,112]. Some other studies [198] have demonstrated that hyaluronic acid is favorable for cell response, by maintaining chondrogenic phenotype and increasing collagen type II production and angiogenesis during in vivo assays. Another example of an excellent natural polymer is chitosan. Chitosan can easily form polyelectrolyte complexes with hyaluronan and chondroitin sulfate [112,209]. Hu X. et al. [198] tried

to mimic the natural cartilage extracellular matrix by synthesizing a biological hydrogel made of hyaluronic acid, chondroitin sulfate modified with 11-azido-3,6,9-tri-oxaundeca-1-amin and gelatin modified with propiolic acid, via click chemistry. Even though the molecular modifications made to the biopolymers let biological hydrogels have good response (making chondrocytes adhere and proliferate on them during in vitro assays), degradation process was too fast. They showed a loss of 45% w in 4 weeks, and a release of 20% w of gelatin and 10% w of chondroitin sulfate during the first two weeks, leading to macroscopic shrinkage of hydrogels.

Furthermore, when combining both naturally-derived and synthetically-derived polymers, adequate degradation kinetics and biological response can be achieved. Park H. et al. [129] created, by photocrosslinking, injectable hydrogels consisting of methacrylated glycol chitosan (MeGC) and hyaluronic acid. The photopolymerized hydrogels were cytocompatible. The incorporation of hyaluronic acid increased cell proliferation while encapsulated chondrocytes improved the cartilaginous extracellular matrix production. Following the same guideline, based on fabricating a hybrid scaffold containing both biological and synthetic components, B.R. Mintz et al. [209] studied a hybrid scaffold, made of a hyaluronic acid-based hydrogel combined with a porous poly(ϵ -caprolactone) material. They tried to understand better the interface and the potential for integration between tissue engineered cartilage scaffolds and surrounding native tissue. They noticed that precursors provide a microenvironment which supports chondrocyte infiltration and proliferation, while maintaining seeded phenotype and structural integrity over a 6-week culture period.

4. Future Trends: From Combination of Materials to Hybrid Hydrogels

Identified problems hindering the application of hydrogels in cartilage regeneration, as described along this paper, include: mechanical properties [210] and mechanical instability [209,211]; dedifferentiation of chondrocytes [163]; toxicity of some of the used crosslinking agents [180]; poor control of gelation kinetics [188,189]; unsuitable degradation kinetics [190–194]. Figure 7 illustrates the combining requirements needed to create materials with biomimetic features.

Mechanical instability inhibits the integration of hydrogels with the surrounding native cartilage tissue when they are implanted [209,211]. In order to synthesize mechanically stable hydrogels and improve their mechanical properties, several options have been proposed. One of the most promising options relays on the principle that materials combination must show the ability to support matrix formation [210], as demonstrated by Boere K.W.M et al. [80]. In this research, it was determined that, when grafting two materials covalently (a 3-D-fabricated poly(hydroxymethylglycolide-co- ϵ -caprolactone)/poly(ϵ -caprolactone) thermoplastic polymer scaffold, functionalized with methacrylate groups and covalently linked to a chondrocyte-laden gelatin methacrylamide hydrogel), the binding strength between the materials improved significantly, resulting in the enhanced mechanical integrity of the reinforced hydrogel. Embedded chondrocytes in hydrogel scaffolds also showed significant cartilage-specific matrix deposition, both in vitro and in vivo assays.

Another promising option to enhance mechanical stability is by regenerating cartilage and bone tissue simultaneously using a two-phased scaffold, since ceramic-to-bone interface has a better and faster integration compared to hydrogel-to-cartilage interface [80]. Additionally it has been observed that bone integration is much faster than cartilage integration, occurring during 2 and 24 weeks after transplantation, respectively.

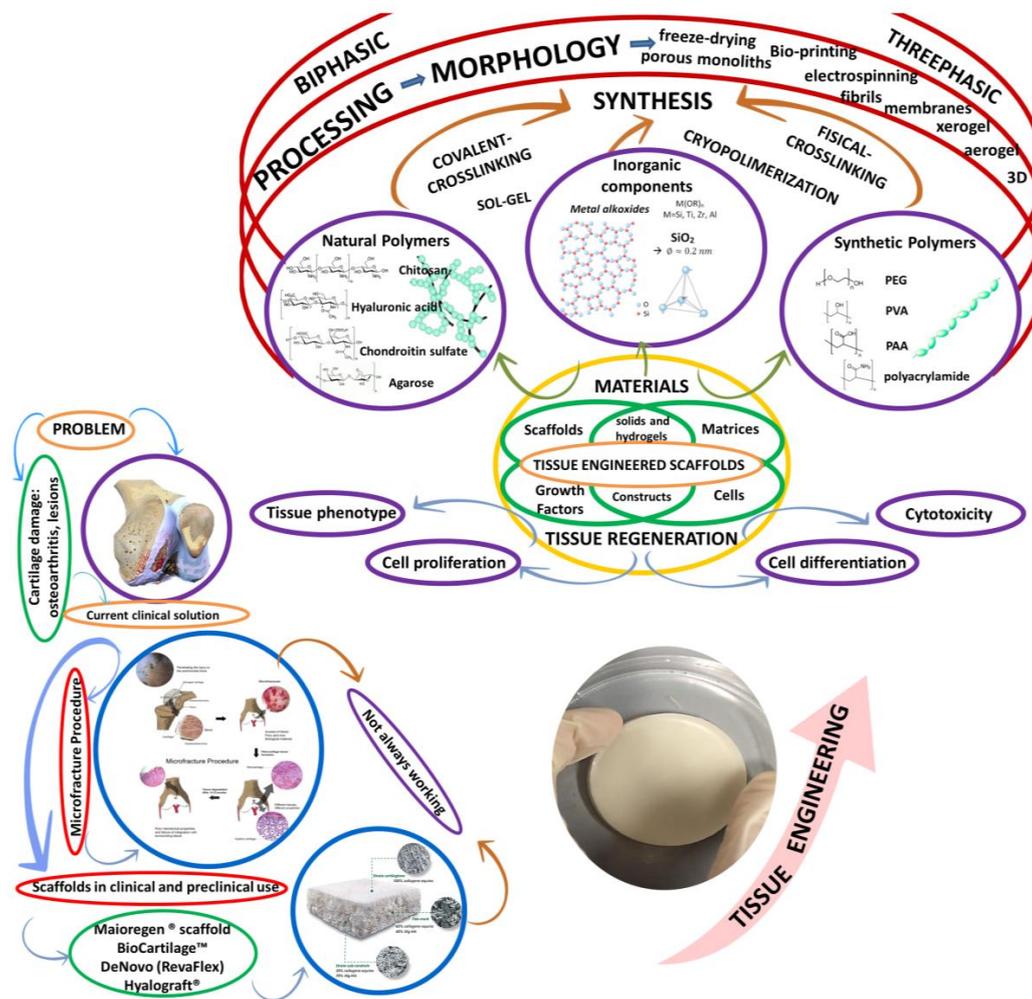


Figure 7. Correlation between current cell-based scaffolds used for cartilage regeneration. Original illustration designed and provided by the authors.

It is possible to have a stable fixation of a cartilage scaffold by exploring a fixation technique with the subchondral bone [206]. One way to accomplish this stable fixation is fabricating an osteochondral scaffold which facilitates fixation and integration with the surrounding cartilage tissue, accelerating the repair of defected articular cartilage when implanted. The general idea is that bone scaffolds act as anchors, providing mechanical stability for cartilage tissue regeneration, besides, the joint between the bone component and the cartilage component should be strong enough to prevent dislocation or delamination on in vivo environment. In order to follow the theory mentioned above, Seol Y.-J. et al. [206] reinforced osteochondral scaffolds by developing combined scaffolds, made of hydrogel scaffolds anchoring to cartilage tissue and ceramic scaffolds anchoring to bone tissue. For in vivo assay, the combined scaffolds were press-fitted into osteochondral tissue defects, in rabbit knee joints. Hydrogel scaffolds and combined scaffolds were compared. After 12 weeks, in vivo experiments demonstrated that regeneration of osteochondral tissue, especially articular cartilage tissue regeneration, was better with combined scaffolds than with hydrogel scaffolds. Hydrogel scaffolds could not keep their initial position, suggesting that ceramic scaffolds in combined scaffolds provided mechanical stability for hydrogel scaffolds. Moreover, G. Camci-Unal et al. [202] realized that combined hydrogels can be biologically and physically tuned to yield within a range of different cell responses and, according to these responses, combined hydrogels may show potential therapeutic possibilities to treat either chondral or osteochondral lesions.

Following this trend, Yang S.S. et al. [138] developed a 3D plotting system to manufacture a biphasic graft which consists of cartilage and subchondral bone for application to osteochondral defects. A combined material (PLGH/alginate) was fabricated as supporting structure to induce a mature osteochondral graft. Cartilage-derived ECM or hydroxyapatite substances were blended with alginate and plotted together with human fetal cartilage-derived progenitor cells, either in the cartilage layer or in the subchondral bone one. The plotted biphasic osteochondral graft showed good integration between layers because no structural separation was observed, while there was dominant cartilage and bone tissue formation during differentiation assay. One of the limitations of using the osteochondral approach in combination with bone marrow derived MSCs is their terminal differentiation, as they seem to follow an endochondral ossification which can arrest differentiation at a stable cartilage hyaline-like phenotype during the chondrogenic process. For this reason, a chondrogenic stimulator, such as the recently described kartogenin which regulates Runx1 expression [212], has to be incorporated with a known inducer of chondrogenic differentiation and a suppressor of hypertrophy.

Another alternative to fulfill the inadequate mechanical strength of hydrogel is constructing a solid-supported thermogel, comprising hydrogel systems or demineralized bone matrix. Huang H. et al. [213] combined chitosan thermogel with demineralized bone matrix to produce solid-supported hydrogel scaffolds. This type of scaffolds provided sufficient strength for cartilage regeneration. They retained homogeneously more bone-derived mesenchymal stem cells (BMSCs) and they proved to have superior matrix production and chondrogenic differentiation in comparison with pure hydrogels and demineralized matrix by their own.

Using fibers of different natural materials to reinforce hydrogels is another way to offer mechanical strength to hybrid scaffolds [205,211]. Mechanical characteristics can be improved or modified using different strategies: (1) varying the number of fiber layers in the laminate; (2) combining different kinds of fibers and nanofiber sheets; (3) modifying crosslinking degree of hydrogels and fibers; (4) changing fibers content and surface treatment; (5) shifting fiber orientation. Fibers anisotropy is an excellent property to reach strong mechanical reinforcement at low charge levels. For example, Buchtová N. et al. [214] developed an injectable hydrogel based on a silanized cellulose derivative: hydroxypropyl methylcellulose interlinked with silica fibers. They proved that the compressive modulus of the hydrogel could be tunable, depending on the covalent bonding between biopolymer and silica fibers. In other approaches, the incorporation of bioactive species, such as cells, growth factors, peptides and proteins into the materials, is proposed to improve hydrogel scaffolds properties [73,215].

It can be deduced, from the reviewed studies, that combination of components yield up reinforced mechanical properties. However, to overcome the other requirements mentioned above, something different has to be done from the already investigated methods to fabricate materials. In authors' opinion, there is a barely explored alternative in material science: studying the potential of hybrid hydrogels based on natural polymers and inorganic components. Hybrids are considered to be materials formed by two components bonded at a molecular level. Commonly one of these components is organic and the other one is inorganic. The new hybrid materials (Figure 8) may show superior characteristics in comparison with the two component phases. This possibility may offer a great potential to design new materials with the complex properties required for cartilage regeneration.

Since degradation rate and mechanical properties can be fine-tuned through chemical and/or physical modifications on either naturally or synthetically derived scaffolds, an excellent opportunity is given to hybrids to emerge as a promising solution for cartilage regeneration. When fine-tuned, hybrids can acquire amorphous, semicrystalline, hydrogen-bonded or supramolecular physical structures [109,210].

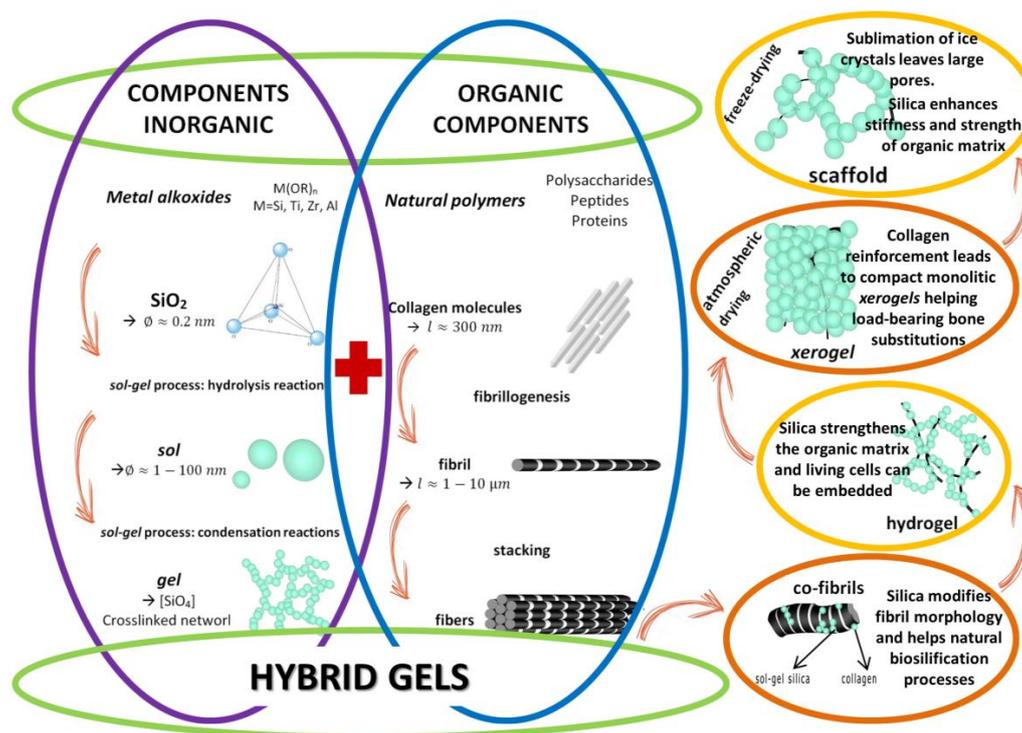


Figure 8. Advantages of hybrid gels. From soft mineralized hydrogels to hard compact xerogels. Original illustration designed and provided by the authors.

Hydrogels made from purely organic precursors are being used as viable materials for cartilage repair due to their ability to retain large portions of water, swollen, distend and exhibit large changes of dimensions (volume changes of several- to 10-fold are common) [216,217]. These characteristics provide hydrogels with a low interfacial tension with water and other fluids, allowing them to reduce mechanical friction between tissues during implantation. The hydrophilic characteristics of hydrogels are caused by the presence of special hydrophilic molecules ($-OH$, $-CONH$, $-CONH_2$, and $-SO_3H$) found in the polymeric components. These molecules give hydrogels different absorption potential [216] and the ability to respond to a range of different stimuli, including temperature, pH, salt, specific (bio)chemical signals, and electric fields [217]. Nevertheless, hydrogels, based on a certain type of precursors, undergo uniform volumetric expansion and contraction, in response to several stimuli; therefore, the great added value of hybrid hydrogels is combining their precursors potential and restrictions. For example, to determine their degradation kinetics and enhanced biological and mechanical properties, it is possible to use: stiffer components like silica-based materials, which restrict swelling in hydrogels in certain positions or directions; or natural polysaccharide-based polymers hybridized with inorganic materials (hydroxyapatite, SiO_2 , or demineralized bone matrix), which restrict the number and type of hydrophilic molecules.

A further step may be the integration of nanotubes with different chemical composition into hybrid scaffolds which may provide them with bioactive and mechanical properties. This is the case of three-dimensional porous collagen sponges where single-walled carbon nanotubes are incorporated into [218]. The incorporation of single-walled carbon nanotubes improved cell proliferation and GAGs production in the *in vivo* microenvironment, because nanotubes were internalized by cells, with benefit for controlled and localized delivery of biological factors. Another study is the design of a biomimetic nanostructured composite cartilage scaffold, via biologically-inspired rosette nanotubes (RNTs) and biocompatible non-woven poly (L-lactic acid) (PLLA) [219]. It was concluded that, RNTs have a similar morphology with native collagen fibers when self-assembled in aqueous conditions, and besides they increase glycosaminoglycan, collagen and protein production; their nanotopography and

surface chemistry enhance chondrogenic differentiation. Another study tested the biocompatibility of 3D artificial hexagonal-pore shaped hybrid organic-inorganic microstructured scaffolds in a rabbit model [220].

The association of silica and polysaccharides within composites or hybrids has demonstrated therapeutic benefits in a wide range of bio-inspired silica-collagen materials [221]. Although this kind of materials has been prepared over nearly 15 years, their application in cartilage regeneration treatments has not been exploited. The great value of hybrid materials (Figure 7) is that it can be synthesized a large variety of structures and properties, from soft mineralized hydrogels to hard compact xerogel, depending on the soft (cartilage) or hard (osteocondral) tissue wanted to be repaired or regenerated. Moreover, to fully comprehend these potential materials and to raise their value for the development of innovative biomedical devices for cartilage regeneration, it is important studying the interplay between the organic-inorganic precursors, which is to follow carefully the polymer self-assembly process and the inorganic condensation mechanisms. Therefore, biological, mechanical and degradation properties can be modulated, guaranteeing bioactivity, cytocompatibility, and an eventual biocompatibility.

5. Concluding Remarks

Currently, there are no clinical satisfactory solutions for cartilage tissue regeneration. From this problem, it raises the need for developing new types of materials and designing more suitable scaffolds which can be used for cartilage regeneration. Investigations based on cell-free hydrogel scaffolds have focused on the optimization of physical-chemical and mechanical properties of matrices. Whereas, cell-seeded hydrogel scaffold studies focus on whether or not they provide an appropriate biochemical and biomechanical environment for regenerating a long-lasting hyaline-type cartilage. Since many materials neither exhibit a low friction coefficient nor withstand several loading cycles, many combinations of polysaccharides and synthetic hydrogels have been assayed to obtain load-bearing and tribological properties similar to native cartilage tissue ones. However, some unsolved problems hinder the application of these materials to the clinic: (1) toxicity of some crosslinking agents; (2) lack of mechanical integrity; (3) poor control of gelation kinetics; (4) unsuitable degradation kinetics.

Among the most promising options, to synthesize mechanically stable hydrogels which support matrix formation, there is the combination of materials to regenerate cartilage and bone tissue simultaneously, using a two/several-phased scaffold. These combinations can be biologically and physically tuned to yield, within a range of different cell responses and according to these responses, combined hydrogels which may show potential therapeutic possibilities to treat either chondral or osteochondral lesions. Osteochondral treatment introduces a new problem and requires the use of a chondrogenic stimulator, such as the recently described kartogenin, to induce chondrogenic differentiation and suppress hypertrophy. From materials science perspective, hybrid hydrogels based on natural polymers and inorganic components may offer a fine tuning of mechanical and biological response, required to reproduce the complexity of the cartilage tissue environment. Silica, hydroxyapatite or demineralized bone matrix/polysaccharide-based hybrids restrict swelling in hydrogels in certain positions or directions by reducing the number and the type of hydrophilic molecules. The association of silica and polysaccharides within composites or hybrids has demonstrated therapeutic benefits in a wide range of bio-inspired silica-collagen materials. To fully comprehend these potential materials and to raise their value for the development of innovative biomedical devices for cartilage regeneration, it is important studying the interplay between the organic-inorganic precursors, which is to follow carefully the polymer self-assembly process and the inorganic condensation mechanisms. Therefore, biological, mechanical and degradation properties can be modulated, bioactivity and cytocompatibility guaranteed and biocompatibility eventually achieved.

Acknowledgments: This work was supported by SIP-IPN 20170510 Project (Mexico), DGICYT Project, MAT2014-51918-C2-I-R (Spain), (CONACYT/CSIC)-EMHE program through project No. MHE200011 (Mexico-Spain). DA Sánchez-Téllez also acknowledges CONACYT (Mexico) for the scholarship given. The authors acknowledge Stephanie Cisneros-Téllez for helping them in the digitalization of Figures 1, 4 and 8.

Author Contributions: Daniela Anahí Sánchez-Téllez, Lucía Téllez-Jurado and Luís María Rodríguez-Lorenzo gave substantial contributions to the conception and design of the review content. Daniela Anahí Sánchez-Téllez wrote the paper. Daniela Anahí Sánchez-Téllez and Luís María Rodríguez-Lorenzo critically revised the final manuscript, making improvements in the drafting of the work. Daniela Anahí Sánchez-Téllez, Daniela Anahí Sánchez-Téllez and Luís María Rodríguez-Lorenzo gave final approval of the version to be published. The three authors, Daniela Anahí Sánchez-Téllez, Lucía Téllez-Jurado and Luís María Rodríguez-Lorenzo agreed in all aspects of the final written work, ensuring that any question related to the accuracy or integrity of any part of the work was appropriately investigated and resolved.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Fickert, S.; Schattenberg, T.; Niks, M.; Weiss, C.; Their, S. Feasibility of arthroscopic 3-dimensional, purely autologous chondrocyte transplantation for chondral defects of the hip: A case series. *Arch. Orthop. Trauma Surg.* **2014**, *134*, 971–978. [[CrossRef](#)] [[PubMed](#)]
2. Baynat, C.; Andro, C.; Vincent, J.P.; Schiele, P.; Buisson, P.; Dubrana, F.; Gunepin, F.X. Actifit[®] synthetic meniscal substitute: Experience with 18 patients in Brest, France. *Orthop. Traumatol. Surg. Res.* **2014**, *100* (Suppl. 8), S385–S389. [[CrossRef](#)] [[PubMed](#)]
3. Verdonk, P.; Dhollander, A.; Almqvist, K.F.; Verdonk, R.; Victor, J. Treatment of osteochondral lesions in the knee using a cell-free scaffold. *Bone Jt. J.* **2015**, *97B*, 318–323. [[CrossRef](#)] [[PubMed](#)]
4. Case, J.M.; Scopp, J.M. Treatment of articular cartilage defects of the knee with microfracture and enhanced microfracture techniques. *Sports Med. Arthrosc.* **2016**, *24*, 63–68. [[CrossRef](#)] [[PubMed](#)]
5. Erggelet, C. Enhanced marrow stimulation techniques for cartilage repair. *Oper. Tech. Orthop.* **2014**, *24*, 2–13. [[CrossRef](#)]
6. Gille, J.; Behrens, P.; Volpi, P.; de Girolamo, L.; Reiss, E.; Zoch, W.; Anders, S. Outcome of autologous matrix induced chondrogenesis (AMIC) in cartilage knee surgery: Data of the AMIC registry. *Arch. Orthop. Trauma Surg.* **2013**, *133*, 87–93. [[CrossRef](#)] [[PubMed](#)]
7. Turajane, T.; Thitiset, T.; Honsawek, S.; Chaveewanakorn, U.; Aojanepong, J.; Papadopoulos, K.I. Assessment of chondrogenic differentiation potential of autologous activated peripheral blood stem cells on human early osteoarthritic cancellous tibial bone scaffold. *Musculoskelet. Surg.* **2014**, *98*, 35–43. [[CrossRef](#)] [[PubMed](#)]
8. Gaharwar, A.K.; Schexnailder, P.J.; Shmidt, G. Chapter 24: Nanocomposite polymer biomaterials for tissue repair of bone and cartilage: A material science perspective. In *Nanomaterials Handbook*; Taylor and Francis Group, LLC: Boca Raton, FL, USA, 2011; pp. 1–20.
9. Chung, C.; Burdick, J.A. Engineering cartilage tissue. *Adv. Drug Deliv. Rev.* **2008**, *60*, 243–262. [[CrossRef](#)] [[PubMed](#)]
10. Chang, G.; Xia, C.; Sherman, O.; Strauss, E.; Jazrawi, L.; Recht, M.P.; Regatte, R.R. High resolution morphologic imaging and T2 mapping of cartilage at 7 tesla: Comparison of cartilage repair patients and healthy controls. *Magn. Reson. Mater. Phys.* **2013**, *26*, 539–548. [[CrossRef](#)] [[PubMed](#)]
11. Mathieu, C.; Chevrier, A.; Lascau-Coman, V.; Rivard, G.E.; Hoemann, C.D. Stereological analysis of subchondral angiogenesis induced by chitosan and coagulation factors in microdrilled articular cartilage defects. *Osteoarthr. Cartil.* **2013**, *21*, 849–859. [[CrossRef](#)] [[PubMed](#)]
12. Siclari, A.; Mascaró, G.; Gentili, C.; Kaps, C.; Cancedda, R.; Boux, E. Cartilage repair in the knee with subchondral drilling augmented with a platelet-rich plasma-immersed polymer-based implant. *Knee Surg. Sports Traumatol. Arthrosc.* **2014**, *22*, 1225–1234. [[CrossRef](#)] [[PubMed](#)]
13. Accardi, M.A.; McCullen, S.D.; Callanan, A.; Chung, S.; Cann, P.M.; Stevens, M.M.; Dini, D. Effects of fiber orientation on the frictional properties and damage of regenerative articular cartilage surfaces. *Tissue Eng. Part A* **2013**, *19*, 2300–2310. [[CrossRef](#)] [[PubMed](#)]
14. Lebourg, M.; Rochina, J.R.; Sousa, T.; Mano, J.; Ribelles, J.L. Different hyaluronic acid morphology modulates primary articular chondrocyte behavior in hyaluronic acid-coated polycaprolactone scaffolds. *J. Biomed. Mater. Res. Part A* **2013**, *101A*, 518–527. [[CrossRef](#)] [[PubMed](#)]

15. Levorson, E.J.; Hu, O.; Mountziaris, P.M.; Kasper, F.K.; Mikos, A.G. Cell-derived polymer/extracellular matrix composite scaffolds for cartilage regeneration, Part 2: Construct devitalization and determination of chondroinductive capacity. *Tissue Eng. Part C Methods* **2014**, *20*, 358–372. [[CrossRef](#)] [[PubMed](#)]
16. Levingstone, T.J.; Matsiko, A.; Dickson, G.R.; O'Brien, F.J.; Gleeson, J.P. A biomimetic multi-layered collagen-based scaffold for osteochondral repair. *Acta Biomater.* **2014**, *10*, 1996–2004. [[CrossRef](#)] [[PubMed](#)]
17. Sutherland, A.J.; Beck, E.C.; Dennis, S.C.; Converse, G.L.; Hopkins, R.A.; Berkland, C.J.; Detamore, M.S. Decellularized cartilage may be a chondroinductive material for osteochondral tissue engineering. *PLoS ONE* **2015**, *10*, e0121966. [[CrossRef](#)] [[PubMed](#)]
18. Sancho-Tello, M.; Forriol, F.; Gastaldi, P.; Ruiz-Saurí, A.; Martín de Llano, J.J.; Novella-Maestre, E.; Antolinos-Turpín, C.M.; Gómez-Tejedor, J.A.; Gómez Ribelles, J.L.; Carda, C. Time evolution of in vivo articular cartilage repair induced by bone marrow stimulation and scaffold implantation in rabbits. *Int. J. Artif. Organs* **2015**, *38*, 210–223. [[CrossRef](#)] [[PubMed](#)]
19. Wang, J.; Yang, Q.; Cheng, N.; Tao, X.; Zhang, Z.; Sun, X.; Zhang, Q. Collagen/silk fibroin composite scaffold incorporated with PLGA microsphere for cartilage repair. *Mater. Sci. Eng. C* **2016**, *61*, 705–711. [[CrossRef](#)] [[PubMed](#)]
20. Shao, Z.; Zhang, X.; Pi, Y.; Yin, L.; Li, L.; Chen, H.; Zhou, C.; Ao, Y. Surface modification on polycaprolactone electrospun mesh and human decalcified bone scaffold with synovium-derived mesenchymal stem cells-affinity peptide for tissue engineering. *J. Biomed. Mater. Res. Part A* **2015**, *103A*, 318–329. [[CrossRef](#)] [[PubMed](#)]
21. Zhang, Y.; Tang, C.L.; Chen, W.J.; Zhang, Q.; Wang, S.L. Dynamic compression combined with exogenous SOX-9 promotes chondrogenesis of adipose-derived mesenchymal stem cells in PLGA scaffold. *Eur. Rev. Med. Pharmacol. Sci.* **2015**, *19*, 2671–2678. [[PubMed](#)]
22. Lehmann, M.; Martin, F.; Mannigel, K.; Kaltschmidt, K.; Sack, U.; Anderer, U. Three-dimensional scaffold-free fusion culture: The way to enhanced chondrogenesis of in vitro propagated human articular chondrocytes. *Eur. J. Histochem.* **2013**, *57*, e31. [[CrossRef](#)] [[PubMed](#)]
23. Kwon, H.; Sun, L.; Cairns, D.M.; Rainbow, R.S.; Preda, R.C.; Kaplan, D.L.; Zeng, L. The influence of scaffold material on chondrocytes under inflammatory conditions. *Acta Biomater.* **2013**, *9*, 6563–6575. [[CrossRef](#)] [[PubMed](#)]
24. Pretzel, D.; Linss, S.; Ahrem, H.; Endres, M.; Kaps, C.; Klemm, D.; Kinne, R.W. A novel in vitro bovine cartilage punch model for assessing the regeneration of focal cartilage defects with biocompatible bacterial nanocellulose. *Arthritis Res. Ther.* **2013**, *15*, R59. [[CrossRef](#)] [[PubMed](#)]
25. Chomchalao, P.; Pongcharoen, S.; Sutheerawattananonda, M.; Tiyaonchai, W. Fibroin and fibroin blended three-dimensional scaffolds for rat chondrocyte culture. *Biomed. Eng. Online* **2013**, *12*, 28. [[CrossRef](#)] [[PubMed](#)]
26. Lee, S.U.; Lee, J.Y.; Joo, S.Y.; Lee, Y.S.; Jeong, C. Transplantation of a scaffold-free cartilage tissue analogue for the treatment of physeal cartilage injury of the proximal tibia in rabbits. *Yonsei Med. J.* **2016**, *57*, 441–448. [[CrossRef](#)] [[PubMed](#)]
27. O'Sullivan, N.A.; Kobayashi, S.; Ranka, M.P.; Zaleski, K.L.; Yaremchuk, M.J.; Bonassar, L.J.; Randolph, M.A. Adhesion and integration of tissue engineered cartilage to porous polyethylene for composite ear reconstruction. *J. Biomed. Mater. Res. Part B* **2015**, *103B*, 983–991. [[CrossRef](#)] [[PubMed](#)]
28. Schleicher, I.; Lips, K.S.; Sommer, U.; Schappat, I.; Martin, A.P.; Szalay, G.; Schnettler, R. Allogeneous bone with collagen for repair of deep osteochondral defects. *J. Surg. Res.* **2013**, *185*, 667–675. [[CrossRef](#)] [[PubMed](#)]
29. Chang, N.J.; Lam, C.F.; Lin, C.C.; Chen, W.L.; Li, C.F.; Lin, Y.T.; Yeh, M.L. Transplantation of autologous endothelial progenitor cells in porous PLGA scaffolds create a microenvironment for the regeneration of hyaline cartilage in rabbits. *Osteoarthr. Cartil.* **2013**, *21*, 1613–1622. [[CrossRef](#)] [[PubMed](#)]
30. Sharma, S.; Lee, A.; Choi, K.; Kim, K.; Youn, I.; Trippel, S.B.; Panitch, A. Biomimetic aggrecan reduces cartilage extracellular matrix from degradation and lowers catabolic activity in ex vivo and in vivo models. *Macromol. Biosci.* **2013**, *13*, 1228–1237. [[CrossRef](#)] [[PubMed](#)]
31. Ding, C.; Qiao, Z.; Jiang, W.; Li, H.; Wei, J.; Zhou, G.; Dai, K. Regeneration of a goat femoral head using a tissue-specific, biphasic scaffold fabricated with CAD/CAM technology. *Biomaterials* **2013**, *34*, 6706–6716. [[CrossRef](#)] [[PubMed](#)]

32. Waldorff, E.I.; Roessler, B.J.; Zachos, T.A.; Miller, B.S.; McHugh, J.; Goldstein, S.A. Preclinical evaluation of a novel implant for treatment of a full-thickness distal femoral focal cartilage defect. *J. Arthroplast.* **2013**, *28*, 1421–1429. [[CrossRef](#)] [[PubMed](#)]
33. Camarero-Espinosa, S.; Rothen-Rutishauser, B.; Weder, C.; Foster, E.J. Directed cell growth in multi-zonal scaffolds for cartilage tissue engineering. *Biomaterials* **2016**, *74*, 42–52. [[CrossRef](#)] [[PubMed](#)]
34. Krych, A.J.; Wanivenhaus, F.; Ng, K.W.; Doty, S.; Warren, R.F.; Maher, S.A. Matrix generation within a macroporous non-degradable implant for osteochondral defects is not enhanced with partial enzymatic digestion of the surrounding tissue: Evaluation in an in vivo rabbit model. *J. Mater. Sci. Mater. Med.* **2013**, *24*, 2429–2437. [[CrossRef](#)] [[PubMed](#)]
35. Irion, V.H.; Flanigan, D.C. New and emerging techniques in cartilage repair: Other scaffold-based cartilage treatment options. *Oper. Tech. Sports Med.* **2013**, *21*, 125–137. [[CrossRef](#)]
36. Kim, Y.S.; Park, D.Y.; Cho, Y.H.; Chang, J.W.; Choi, J.W.; Park, J.K.; Min, B.H.; Shin, Y.S.; Kim, C.H. Cultured chondrocyte and porcine cartilage-derived substance (PCS) construct as a possible dorsal augmentation material in rhinoplasty: A preliminary animal study. *J. Plast. Reconstr. Aesthet. Surg.* **2015**, *68*, 659–666. [[CrossRef](#)] [[PubMed](#)]
37. Filardo, G.; Kon, E.; Perdisa, F.; Di Matteo, B.; Di Martino, A.; Iacono, F.; Zaffagnini, S.; Balboni, F.; Vaccari, V.; Marcacci, M. Osteochondral scaffold reconstruction for complex knee lesions: A comparative evaluation. *Knee* **2013**, *20*, 570–576. [[CrossRef](#)] [[PubMed](#)]
38. Kon, E.; Filardo, G.; Di Martino, A.; Busacca, M.; Moio, A.; Perdisa, F.; Marcacci, M. Clinical results and mri evolution of a nano-composite multilayered biomaterial for osteochondral regeneration at 5 years. *Am. J. Sports Med.* **2014**, *42*, 158–165. [[CrossRef](#)] [[PubMed](#)]
39. Kon, E.; Filardo, G.; Perdisa, F.; Di Martino, A.; Busacca, M.; Balboni, F.; Sessa, A.; Marcacci, M. A one-step treatment for chondral and osteochondral knee defects: Clinical results of a biomimetic scaffold implantation at 2 years of follow-up. *J. Mater. Sci. Mater. Med.* **2014**, *25*, 2437–2444. [[CrossRef](#)] [[PubMed](#)]
40. Caminal, M.; Moll, X.; Codina, D.; Rabanal, R.M.; Morist, A.; Barrachina, J.; Garcia, F.; Pla, A.; Vives, J. Transitory improvement of articular cartilage characteristics after implantation of polylactide: Polyglycolic acid (PLGA) scaffolds seeded with autologous mesenchymal stromal cells in a sheep model of critical-sized chondral defect. *Biotechnol. Lett.* **2014**, *36*, 2143–2153. [[CrossRef](#)] [[PubMed](#)]
41. Delcogliano, M.; de Caro, F.; Scaravella, E.; Ziveri, G.; De Biase, C.F.; Marotta, D.; Marengi, P.; Delcogliano, A. Use of innovative biomimetic scaffold in the treatment for large osteochondral lesions of the knee. *Knee Surg. Sports Traumatol. Arthrosc.* **2014**, *22*, 1260–1269. [[CrossRef](#)] [[PubMed](#)]
42. McCormick, F.; Cole, B.J.; Nwachukwu, B.; Harris, J.D.; Adkisson, H.D.; Farr, J. Treatment of focal cartilage defects with a juvenile allogeneic 3-dimensional articular cartilage graft. *Oper. Techniq. Sports Med.* **2013**, *21*, 95–99. [[CrossRef](#)]
43. Chiang, H.; Liao, C.J.; Hsieh, C.H.; Shen, C.Y.; Huang, Y.Y.; Jiang, C.C. Clinical feasibility of a novel biphasic osteochondral composite for matrix-associated autologous chondrocyte implantation. *Osteoarthr. Cartil.* **2013**, *21*, 589–598. [[CrossRef](#)] [[PubMed](#)]
44. Kon, E.; Filardo, G.; Venieri, G.; Perdisa, F.; Marcacci, M. Tibial plateau lesions. Surface reconstruction with a biomimetic osteochondral scaffold: Results at 2 years of follow-up. *Inj. Int. J. Care Inj.* **2014**, *45S*, S121–S125. [[CrossRef](#)] [[PubMed](#)]
45. Abrams, G.A.; Mall, N.A.; Fortier, L.A.; Roller, B.L.; Cole, B.J. BioCartilage: Background and operative technique. *Oper. Tech. Sports Med.* **2013**, *21*, 116–124. [[CrossRef](#)]
46. Sharma, B.; Fermanian, S.; Gibson, M.; Unterman, S.; Herzka, D.A.; Cascio, B.; Coburn, J.; Hui, A.Y.; Marcus, N.; Gold, G.E.; et al. Human cartilage repair with a photoreactive adhesive-hydrogel composite. *Sci. Transl. Med.* **2013**, *5*, 167ra6. [[CrossRef](#)] [[PubMed](#)]
47. Quarch, V.M.; Enderle, E.; Lotz, J.; Frosch, K.H. Fate of large donor site defects in osteochondral transfer procedures in the knee joint with and without TruFit plugs. *Arch. Orthop. Trauma Surg.* **2014**, *134*, 657–666. [[CrossRef](#)] [[PubMed](#)]
48. Bartha, L.; Hamann, D.; Pieper, J.; Péters, F.; Riesle, J.; Vajda, A.; Novak, P.K.; Hangody, L.R.; Vasarhelyi, G.; Bodó, L.; et al. A clinical feasibility study to evaluate the safety and efficacy of PEOT/PBT implants for human donor site filling during mosaicplasty. *Eur. J. Orthop. Surg. Traumatol.* **2013**, *23*, 81–91. [[CrossRef](#)] [[PubMed](#)]

49. Porcellini, G.; Merolla, G.; Campi, F.; Pellegrini, A.; Bodanki, C.S.; Paladini, P. Arthroscopic treatment of early glenohumeral arthritis. *J. Orthop. Traumatol.* **2013**, *14*, 23–29. [[CrossRef](#)] [[PubMed](#)]
50. Krase, A.; Abedian, R.; Steck, E.; Hurschler, C.; Richter, W. BMP activation and WNT-signalling affect biochemistry and functional biomechanical properties of cartilage tissue engineering constructs. *Osteoarthr. Cartil.* **2014**, *22*, 284–292. [[CrossRef](#)] [[PubMed](#)]
51. Panadero, J.A.; Vikingsson, L.V.; Gomez Ribelles, J.L.; Lanceros-Mendez, S.; Sencadas, V. In vitro mechanical fatigue behavior of poly- ϵ -caprolactone macroporous scaffolds for cartilage tissue engineering: Influence of pore filling by a poly(vinyl alcohol) gel. *J. Biomed. Mater. Res. Part B Appl. Biomater.* **2014**, *103*, 1037–1043. [[CrossRef](#)] [[PubMed](#)]
52. Motavalli, M.; Whitney, G.A.; Dennis, J.E.; Mansour, J.M. Investigating a continuous shear strain function for depth-dependent properties of native and tissue engineering cartilage using pixel-size data. *J. Mech. Behav. Biomed. Mater.* **2013**, *28*, 62–70. [[CrossRef](#)] [[PubMed](#)]
53. Hendriks, J.A.; Moroni, L.; Riesle, J.; de Wijn, J.R.; van Blitterswijk, C.A. The effect of scaffold-cell entrapment capacity and physico-chemical properties on cartilage regeneration. *Biomaterials* **2013**, *34*, 4259–4265. [[CrossRef](#)] [[PubMed](#)]
54. Griebel, A.J.; Khoshgoftar, M.; Novak, T.; van Donkelaar, C.C.; Neu, C.P. Direct noninvasive measurement and numerical modeling of depth-dependent strains in layered agarose constructs. *J. Biomech.* **2014**, *47*, 2149–2156. [[CrossRef](#)] [[PubMed](#)]
55. Ahn, H.; Kim, K.J.; Park, S.Y.; Huh, J.E.; Kim, H.J.; Yu, W.R. 3D braid scaffolds for regeneration of articular cartilage. *J. Mech. Behav. Biomed. Mater.* **2014**, *34*, 37–46. [[CrossRef](#)] [[PubMed](#)]
56. MacBarb, R.F.; Chen, A.L.; Hu, J.C.; Athanasiou, K.A. Engineering functional anisotropy in fibrocartilage neotissues. *Biomaterials* **2013**, *34*, 9980–9989. [[CrossRef](#)] [[PubMed](#)]
57. Moradi, A.; Pramanik, S.P.; Ataollahi, F.; Khalil, A.A.; Kamarul, T.; Pinguan-Murphy, B. A comparison study of different physical treatments on cartilage matrix derived porous scaffolds for tissue engineering applications. *Sci. Technol. Adv. Mater.* **2014**, *15*, 065001. [[CrossRef](#)] [[PubMed](#)]
58. Peng, G.; McNary, S.M.; Athanasiou, K.A.; Reddi, A.H. Surface zone articular chondrocytes modulate the bulk and surface mechanical properties of the tissue-engineered cartilage. *Tissue Eng. Part A* **2014**, *20*, 3332–3341. [[CrossRef](#)] [[PubMed](#)]
59. Causin, P.; Sacco, R.; Verri, M. A multiscale approach in the computational modeling of the biophysical environment in artificial cartilage tissue regeneration. *Biomech. Model. Mechanobiol.* **2013**, *12*, 763–780. [[CrossRef](#)] [[PubMed](#)]
60. Da, H.; Jia, S.J.; Meng, G.L.; Cheng, J.H.; Zhou, W.; Xiong, Z.; Mu, Y.J.; Liu, J. The impact of compact layer in biphasic scaffold on osteochondral tissue engineering. *PLoS ONE* **2013**, *8*, e54838. [[CrossRef](#)] [[PubMed](#)]
61. Shimomura, K.; Moriguchi, Y.; Ando, W.; Nansai, R.; Fujie, H.; Hart, D.A.; Gobbi, A.; Kita, K.; Horibe, S.; Shino, K.; et al. Osteochondral repair using a scaffold-free tissue-engineered construct derived from synovial mesenchymal stem cells and a hydroxyapatite-based artificial bone. *Tissue Eng. Part A* **2014**, *20*, 2291–2304. [[CrossRef](#)] [[PubMed](#)]
62. Dabiri, Y.; Li, L.P. Influences of the depth-dependent material inhomogeneity of articular cartilage on the fluid pressurization in the human knee. *Med. Eng. Phys.* **2013**, *35*, 1591–1598. [[CrossRef](#)] [[PubMed](#)]
63. Pierce, D.M.; Ricken, T.; Holzapfel, G.A. Modeling sample/patient-specific structural and diffusional responses of cartilage using DT-MRI. *Int. J. Numer. Methods Biomed. Eng.* **2013**, *29*, 807–821. [[CrossRef](#)] [[PubMed](#)]
64. Chang, D.P.; Guilak, F.; Jay, G.D.; Zauscher, S. Interaction of lubricin with type II collagen surfaces: Adsorption, friction, and normal forces. *J. Biomech.* **2014**, *47*, 659–666. [[CrossRef](#)] [[PubMed](#)]
65. Greene, G.W.; Olszewska, A.; Osterberg, M.; Zhu, H.; Horn, R. A cartilage-inspired lubrication system. *Soft Matter* **2014**, *10*, 374–382. [[CrossRef](#)] [[PubMed](#)]
66. Laurenti, K.C.; de Albuquerque Haach, L.C.; dos Santos, A.R., Jr.; de Almeida Rollo, J.M.D.; Bezerra de Menezes Reiff, R.; Minarelli Gaspar, A.M.; de Moraes Purquerio, B.; Fortulan, C.A. Cartilage reconstruction using self-anchoring implant with functional gradient. *Mater. Res.* **2014**, *17*, 638–649. [[CrossRef](#)]
67. Utschneider, S.; Lorber, V.; Dedic, M.; Paulus, A.C.; Schröder, C.; Gottschalk, O.; Schmitt-Sody, M.; Jansson, V. Biological activity and migration of wear particles in the knee joint: An in vivo comparison of six different polyethylene materials. *J. Mater. Sci. Mater. Med.* **2014**, *25*, 1599–1612. [[CrossRef](#)] [[PubMed](#)]

68. Nürnberger, S.; Meyer, C.; Ponomarev, I.; Barnewitz, D.; Resinger, C.; Klepal, W.; Albrecht, C.; Marlovits, S. Equine articular chondrocytes on MACT scaffolds for cartilage defect treatment. *Anat. Histol. Embryol.* **2013**, *42*, 332–343. [[CrossRef](#)] [[PubMed](#)]
69. Vahdati, A.; Wagner, D.R. Implant size and mechanical properties influence the failure of the adhesive bond between cartilage implants and native tissue in a finite element analysis. *J. Biomech.* **2013**, *46*, 1554–1560. [[CrossRef](#)] [[PubMed](#)]
70. Bulman, S.E.; Coleman, C.M.; Murphy, J.M.; Medcalf, N.; Ryan, A.E.; Barry, F. Pullulan: A new cytoadhesive for cell-mediated cartilage repair. *Stem Cell Res. Ther.* **2015**, *6*, 34. [[CrossRef](#)] [[PubMed](#)]
71. Vikingsson, L.; Gómez-Tejedor, J.A.; Gallego Ferrer, G.; Gómez Ribelles, J.L. An experimental fatigue study of a porous scaffold for the regeneration of articular cartilage. *J. Biomech.* **2015**, *48*, 1310–1317. [[CrossRef](#)] [[PubMed](#)]
72. Kharkar, P.M.; Kiick, K.L.; Kloxin, A.M. Designing degradable hydrogels for orthogonal control of cell microenvironments. *Chem. Soc. Rev.* **2013**, *42*, 7335–7372. [[CrossRef](#)] [[PubMed](#)]
73. Delighkaris, K.; Tadele, T.S.; Olthuis, W.; van den Berg, A. Hydrogel-based devices for biomedical applications. *Sens. Actuators B Chem.* **2010**, *147*, 765–774. [[CrossRef](#)]
74. Kim, I.L.; Mauck, R.L.; Burdick, J.A. Hydrogel design for cartilage tissue engineering: A case study with hyaluronic acid. *Biomaterials* **2011**, *32*, 8771–8782. [[CrossRef](#)] [[PubMed](#)]
75. Martins, E.A.N.; Michelacci, Y.M.; Baccarin, R.Y.A.; Cogliati, B.; Silva, L.S.L.C. Evaluation of chitosan-GP hydrogel biocompatibility in osteochondral defects: An experimental approach. *BMC Vet. Res.* **2014**, *10*, 197. [[CrossRef](#)] [[PubMed](#)]
76. Kazusa, H.; Nakasa, T.; Shibuya, H.; Ohkawa, S.; Kamei, G.; Adachi, N.; Deie, M.; Nakajima, N.; Hyon, S.H.; Ochi, M. Strong adhesiveness of a new biodegradable hydrogel glue, LYDEX, for use on articular cartilage. *J. Appl. Biomater. Funct. Mater.* **2013**, *11*, 180–186. [[CrossRef](#)] [[PubMed](#)]
77. Mesallati, T.; Buckley, C.T.; Kelly, D.J. Engineering articular cartilage-like grafts by self-assembly of infrapatellar fat pad-derived stem cells. *Biotechnol. Bioeng.* **2014**, *111*, 1686–1698. [[CrossRef](#)] [[PubMed](#)]
78. Bhat, S.; Lidgren, L.; Kumar, A. In vitro neo-cartilage formation on a three-dimensional composite polymeric cryogel matrix. *Macromol. Biosci.* **2013**, *13*, 827–837. [[CrossRef](#)] [[PubMed](#)]
79. Foss, C.; Merzari, E.; Migliaresi, C.; Motta, A. Silk fibroin/hyaluronic acid 3D matrices for cartilage tissue engineering. *Biomacromolecules* **2013**, *14*, 38–47. [[CrossRef](#)] [[PubMed](#)]
80. Boere, K.W.; Visser, J.; Seyednejad, H.; Rahimian, S.; Gawlitta, D.; van Steenberg, M.J.; Dhert, W.J.; Hennink, W.E.; Vermonden, T.; Malda, J. Covalent attachment of a three-dimensionally printed thermoplast to a gelatin hydrogel for mechanically enhanced cartilage constructs. *Acta Biomater.* **2014**, *10*, 2602–2611. [[CrossRef](#)] [[PubMed](#)]
81. Parmar, P.A.; Chow, L.W.; St-Pierre, J.P.; Horejs, C.M.; Peng, Y.Y.; Werkmeister, J.A.; Ramshaw, J.A.; Stevens, M.M.; Paresch, A.; Parmar, L.W.C.; et al. Collagen-mimetic peptide-modifiable hydrogels for articular cartilage regeneration. *Biomaterials* **2015**, *54*, 213–225. [[CrossRef](#)] [[PubMed](#)]
82. Omobono, M.A.; Zhao, X.; Furlong, M.A.; Kwon, C.H.; Gill, T.J.; Randolph, M.A.; Redmond, R.W. Enhancing the stiffness of collagen hydrogels for delivery of encapsulated chondrocytes to articular lesions for cartilage regeneration. *J. Biomed. Mater. Res. Part A* **2015**, *103A*, 1332–1338. [[CrossRef](#)] [[PubMed](#)]
83. Liao, I.C.; Moutos, F.T.; Estes, B.T.; Zhao, X.; Guilak, F. Composite three-dimensional woven scaffolds with interpenetrating network hydrogels to create functional synthetic articular cartilage. *Adv. Funct. Mater.* **2013**, *23*, 5833–5839. [[CrossRef](#)] [[PubMed](#)]
84. Antonioli, E.; Lobo, A.O.; Ferretti, M.; Cohen, M.; Marciano, F.R.; Corat, E.J.; Trava-Airoldi, V.J. An evaluation of chondrocyte morphology and gene expression on superhydrophilic vertically-aligned multi-walled carbon nanotube films. *Mater. Sci. Eng. C* **2013**, *33*, 641–647. [[CrossRef](#)] [[PubMed](#)]
85. Liu, M.; Ishida, Y.; Ebina, Y.; Sasaki, T.; Hikima, T.; Takata, M.; Aida, T. An anisotropic hydrogel with electrostatic repulsion between cofacially aligned nanosheets. *Nature* **2015**, *517*, 68–72. [[CrossRef](#)] [[PubMed](#)]
86. Leone, G.; Bidini, A.B.; Lamponi, S.; Magnani, A. States of water, surface and rheological characterisation of a new biohydrogel as articular cartilage substitute. *Polym. Adv. Technol.* **2013**, *24*, 824–833. [[CrossRef](#)]
87. Drira, Z.; Yadavalli, V.K. Nanomechanical measurements of polyethyleneglycol hydrogels using atomic force microscopy. *J. Mech. Behav. Biomed. Mater.* **2013**, *18*, 20–28. [[CrossRef](#)] [[PubMed](#)]

88. Ronken, S.; Wirz, D.; Daniels, A.U.; Kurokawa, T.; Gong, J.P.; Arnold, M.P. Double-network acrylamide hydrogel compositions adapted to achieve cartilage-like dynamic stiffness. *Biomech. Model. Mechanobiol.* **2013**, *12*, 243–248. [[CrossRef](#)] [[PubMed](#)]
89. Snyder, T.N.; Madhavan, K.; Intrator, M.; Dregalla, R.C.; Park, D. A fibrin/hyaluronic acid hydrogel for the delivery of mesenchymal stem cells and potential for articular cartilage repair. *J. Biol. Eng.* **2014**, *8*, 10. [[CrossRef](#)] [[PubMed](#)]
90. Ahearne, M.; Liu, Y.; Kelly, D.J. Combining freshly isolated chondroprogenitor cells from the infrapatellar fat pad with a growth factor delivery hydrogel as a putative single stage therapy for articular cartilage repair. *Tissue Eng. Part A* **2014**, *20*, 930–939. [[CrossRef](#)] [[PubMed](#)]
91. Mesallati, T.; Buckley, C.T.; Kelly, D.J. A comparison of self-assembly and hydrogel encapsulation as a means to engineer functional cartilaginous grafts using culture expanded chondrocytes. *Tissue Eng. Part C* **2014**, *20*, 52–63. [[CrossRef](#)] [[PubMed](#)]
92. Ramesh, S.; Rajagopal, K.; Vaikkath, D.; Nair, P.D.; Madhuri, V. Enhanced encapsulation of chondrocytes within a chitosan/ hyaluronic acid hydrogel: A new technique. *Biotechnol. Lett.* **2014**, *36*, 1107–1111. [[CrossRef](#)] [[PubMed](#)]
93. Ha, C.W.; Park, Y.B.; Chung, J.Y.; Park, Y.G. Cartilage repair using composites of human umbilical cord blood-derived mesenchymal stem cells and hyaluronic acid hydrogel in a minipig model. *Stem Cells Transl. Med.* **2015**, *4*, 1–8. [[CrossRef](#)] [[PubMed](#)]
94. Zeng, L.; Chen, X.; Zhang, Q.; Yu, F.; Li, Y.; Yao, Y. Redifferentiation of dedifferentiated chondrocytes in a novel three-dimensional microcavitary hydrogel. *J. Biomed. Mater. Res. Part A* **2015**, *103A*, 1693–1702. [[CrossRef](#)] [[PubMed](#)]
95. Murakami, T.; Sakai, N.; Yamaguchi, T.; Yarimitsu, S.; Nakashima, K.; Sawae, Y.; Suzuki, A. Evaluation of a superior lubrication mechanism with biphasic hydrogels for artificial cartilage. *Tribol. Int.* **2015**, *89*, 19–26. [[CrossRef](#)]
96. Chen, K.; Zhang, D.; Cui, X.; Wang, Q. Research on swing friction lubrication mechanisms and the fluid load support characteristics of PVA–HA composite hydrogel. *Tribol. Int.* **2015**, *90*, 412–419. [[CrossRef](#)]
97. Baykal, D.; Underwood, R.J.; Mansmann, K.; Marcolongo, M.; Kurtz, S.M. Evaluation of friction properties of hydrogels based on a biphasic cartilage model. *J. Mech. Behav. Biomed. Mater.* **2013**, *28*, 263–273. [[CrossRef](#)] [[PubMed](#)]
98. Guo, Y.; Guo, J.; Bai, D.; Wang, H.; Zheng, X.; Guo, W.; Tian, W. Hemiarthroplasty of the shoulder joint using a custom-designed high-density nano-hydroxyapatite/ polyamide prosthesis with a polyvinyl alcohol hydrogel humeral head surface in rabbits. *Artif. Organs* **2014**, *38*, 580–586. [[CrossRef](#)] [[PubMed](#)]
99. Chiang, C.-W.; Chen, W.-C.; Liu, H.-W.; Chen, C.-H. Application of synovial fluid mesenchymal stem cells: Platelet-rich plasma hydrogel for focal cartilage defect. *J. Exp. Clin. Med.* **2014**, *6*, 118–124. [[CrossRef](#)]
100. Sridhar, B.V.; Brock, J.L.; Silver, J.S.; Leight, J.L.; Randolph, M.A.; Anseth, K.S. Development of a cellularly degradable PEG hydrogel to promote articular cartilage extracellular matrix deposition. *Adv. Healthc. Mater.* **2015**, *4*, 702–713. [[CrossRef](#)] [[PubMed](#)]
101. Smeriglio, P.; Lai, J.H.; Dhulipala, L.; Behn, A.W.; Goodman, S.B.; Smith, R.L.; Maloney, W.J.; Yang, F.; Bhutani, N. Comparative potential of juvenile and adult human articular chondrocytes for cartilage tissue formation in three-dimensional biomimetic hydrogels. *Tissue Eng. Part A* **2015**, *21*, 147–155. [[CrossRef](#)] [[PubMed](#)]
102. Zhao, F.; He, W.; Yan, Y.; Zhang, H.; Zhang, G.; Tian, D.; Gao, H. The application of polysaccharide biocomposites to repair cartilage defects. *Int. J. Polym. Sci.* **2014**, *2014*, 654597. [[CrossRef](#)]
103. Zeng, L.; Yao, Y.; Wang, D.A.; Chen, X. Effect of microcavitary alginate hydrogel with different pore sizes on chondrocyte culture for cartilage tissue engineering. *Mater. Sci. Eng. C* **2014**, *34*, 168–175. [[CrossRef](#)] [[PubMed](#)]
104. Rackwitz, L.; Djouad, F.; Janjanin, S.; Nöth, U.; Tuan, R.S. Functional cartilage repair capacity of de-differentiated, chondrocyte and mesenchymal stem cell-laden hydrogels in vitro. *Osteoarthr. Cartil.* **2014**, *22*, 1148–1157. [[CrossRef](#)] [[PubMed](#)]
105. Ponnurangam, S.; O’Connell, G.D.; Chernyshova, I.V.; Wood, K.; Hung, C.T.; Somasundaran, P. Beneficial effects of cerium oxide nanoparticles in development of chondrocyte-seeded hydrogel constructs and cellular response to interleukin insults. *Tissue Eng. Part A* **2014**, *20*, 2908–2919. [[CrossRef](#)] [[PubMed](#)]

106. Dashtdar, H.; Murali, M.R.; Abbas, A.A.; Suhaeb, A.M.; Selvaratnam, L.; Tay, L.X.; Kamarul, T. PVA-chitosan composite hydrogel versus alginate beads as a potential mesenchymal stem cell carrier for the treatment of focal cartilage defects. *Knee Surg. Sports Traumatol. Arthrosc.* **2015**, *23*, 1368–1377. [[CrossRef](#)] [[PubMed](#)]
107. Choi, B.; Kim, S.; Fan, J.; Kowalski, T.; Petrigliano, F.; Evseenko, D.; Lee, M. Covalently conjugated transforming growth factor- β 1 in modular chitosan hydrogels for the effective treatment of articular cartilage defects. *Biomater. Sci.* **2015**, *3*, 742. [[CrossRef](#)] [[PubMed](#)]
108. Puppi, D.; Chiellini, F.; Piras, A.M.; Chiellini, E. Polymeric materials for bone and cartilage repair. *Prog. Polym. Sci.* **2010**, *35*, 403–440. [[CrossRef](#)]
109. Tan, H.; Marra, K.G. Injectable, biodegradable hydrogels for tissue engineering applications. *Materials* **2010**, *3*, 1746–1767. [[CrossRef](#)]
110. Seol, D.; Magonetta, M.J.; Ramakrishnan, P.S.; Kurriger, G.L.; Choe, H.; Jang, K.; Martin, J.A.; Lim, T.H. Biocompatibility and preclinical feasibility tests of a temperature-sensitive hydrogel for the purpose of surgical wound pain control and cartilage repair. *J. Biomed. Mater. Res. Part B* **2013**, *101B*, 1508–1515. [[CrossRef](#)] [[PubMed](#)]
111. Petit, A.; Sandker, M.; Müller, B.; Meyboom, R.; van Midwoud, P.; Bruin, P.; Redout, E.M.; Versluijs-Helder, M.; van der Lest, C.H.; Buwalda, S.J.; et al. Release behavior and intra-articular biocompatibility of celecoxib-loaded acetyl-capped PCLA-PEG-PCLA thermogels. *Biomaterials* **2014**, *35*, 7919–7928. [[CrossRef](#)] [[PubMed](#)]
112. Muzzarelli, R.A.A.; Greco, F.; Busilacchi, A.; Sollazzo, V.; Gigante, A. Chitosan, hyaluronan and chondroitin sulfate in tissue engineering for cartilage regeneration: A review. *Carbohydr. Polym.* **2012**, *89*, 723–739. [[CrossRef](#)] [[PubMed](#)]
113. Tamaddon, M.; Walton, R.S.; Brand, D.D.; Czernuszka, J.T. Characterisation of freeze-dried type ii collagen and chondroitin sulfate scaffolds. *J. Mater. Sci. Mater. Med.* **2013**, *24*, 1153–1165. [[CrossRef](#)] [[PubMed](#)]
114. Park, H.; Lee, K.Y. Cartilage regeneration using biodegradable oxidized alginate/hyaluronate hydrogels. *J. Biomed. Mater. Res. Part A* **2014**, *102A*, 4519–4525. [[CrossRef](#)] [[PubMed](#)]
115. Walker, K.J.; Madihally, S.V. Anisotropic temperature sensitive chitosan-based injectable hydrogels mimicking cartilage matrix. *J. Biomed. Mater. Res. Part B* **2015**, *103B*, 1149–1160. [[CrossRef](#)] [[PubMed](#)]
116. Sheehy, E.J.; Mesallati, T.; Vinardell, T.; Kelly, D.J. Engineering cartilage or endochondral bone: A comparison of different naturally derived hydrogels. *Acta Biomater.* **2015**, *13*, 245–253. [[CrossRef](#)] [[PubMed](#)]
117. Kaderli, S.; Viguier, E.; Watrelot-Virieux, D.; Roger, T.; Gurny, R.; Scapozza, L.; Möller, M.; Boulocher, C.; Jordan, O. Efficacy study of two novel hyaluronic acid-based formulations for viscosupplementation therapy in an early osteoarthrotic rabbit model. *Eur. J. Pharm. Biopharm.* **2015**, *96*, 388–395. [[CrossRef](#)] [[PubMed](#)]
118. Ren, C.D.; Gao, S.; Kurisawa, M.; Ying, J.Y. Cartilage synthesis in hyaluronic acid–tyramine constructs. *J. Mater. Chem. B* **2015**, *3*, 1942. [[CrossRef](#)]
119. Kim, J.; Lin, B.; Kim, S.; Choi, B.; Evseenko, D.; Lee, M. TGF- β 1 conjugated chitosan collagen hydrogels induce chondrogenic differentiation of human synovium-derived stem cells. *J. Biol. Eng.* **2015**, *9*, 1. [[CrossRef](#)] [[PubMed](#)]
120. Jovanović, Z.; Radosavljević, A.; Kačarević-Popović, Z.; Stojkowska, J.; Perić-Grujić, A.; Ristić, M.; Matić, I.Z.; Juranić, Z.D.; Obradović, B.; Mišković-Stanković, V. Bioreactor validation and biocompatibility of Ag/poly(n-vinyl-2-pyrrolidone) hydrogel nanocomposites. *Colloids Surf. B Biointerfaces* **2013**, *105*, 230–235. [[CrossRef](#)] [[PubMed](#)]
121. Dua, R.; Centeno, J.; Ramaswamy, S. Augmentation of engineered cartilage to bone integration using hydroxyapatite. *J. Biomed. Mater. Res. Part B* **2014**, *102 B*, 922–932. [[CrossRef](#)] [[PubMed](#)]
122. Pan, Y.; Shen, Q.; Pan, C.; Wang, J. Compressive mechanical characteristics of multi-layered gradient hydroxyapatite reinforced poly (vinyl alcohol) gel biomaterial. *J. Mater. Sci. Technol.* **2013**, *29*, 551–556. [[CrossRef](#)]
123. Yodmuang, S.; McNamara, S.L.; Nover, A.B.; Mandal, B.B.; Agarwal, M.; Kelly, T.A.; Chao, P.H.; Hung, C.; Kaplan, D.L.; Vunjak-Novakovic, G. Silk microfiber-reinforced silk hydrogel composites for functional cartilage tissue repair. *Acta Biomater.* **2015**, *11*, 27–36. [[CrossRef](#)] [[PubMed](#)]
124. Parkes, M.; Myant, C.; Dini, D.; Cann, P. Tribology-optimised silk protein hydrogels for articular cartilage repair. *Tribol. Int.* **2015**, *89*, 9–18. [[CrossRef](#)]
125. Su, R.S.; Kim, Y.; Liu, J.C. Resilin: Protein-based elastomeric biomaterials. *Acta Biomater.* **2014**, *10*, 1601–1611. [[CrossRef](#)] [[PubMed](#)]

126. Kuo, C.Y.; Chen, C.H.; Hsiao, C.Y.; Chen, J.P. Incorporation of chitosan in biomimetic gelatin/chondroitin-6-sulfate/hyaluronan cryogel for cartilage tissue engineering. *Carbohydr. Polym.* **2015**, *117*, 722–730. [[CrossRef](#)] [[PubMed](#)]
127. Zhao, L.; Gwon, H.J.; Lim, Y.M.; Nho, Y.C.; Kim, S.Y. Hyaluronic acid/chondroitin sulfate-based hydrogel prepared by gamma irradiation technique. *Carbohydr. Polym.* **2014**, *102*, 598–605. [[CrossRef](#)] [[PubMed](#)]
128. Ni, Y.; Tang, Z.; Cao, W.; Lin, H.; Fan, Y.; Guo, L.; Zhang, X. Tough and elastic hydrogel of hyaluronic acid and chondroitin sulfate as potential cell scaffold materials. *Int. J. Biol. Macromol.* **2015**, *74*, 367–375. [[CrossRef](#)] [[PubMed](#)]
129. Park, H.; Choi, B.; Hu, J.; Lee, M. Injectable chitosan hyaluronic acid hydrogels for cartilage tissue engineering. *Acta Biomater.* **2013**, *9*, 4779–4789. [[CrossRef](#)] [[PubMed](#)]
130. Yang, R.; Tan, L.; Cen, L.; Zhang, Z. An injectable scaffold based on crosslinked hyaluronic acid gel for tissue regeneration. *RSC Adv.* **2016**, *6*, 16838. [[CrossRef](#)]
131. Fiorica, C.; Palumbo, F.S.; Pitarresi, G.; Gulino, A.; Agnello, S.; Giammona, G. Injectable in situ forming hydrogels based on natural and synthetic polymers for potential application in cartilage repair. *RSC Adv.* **2015**, *5*, 19715. [[CrossRef](#)]
132. Matsuzaki, T.; Matsushita, T.; Tabata, Y.; Saito, T.; Matsumoto, T.; Nagai, K.; Kuroda, R.; Kurosaka, M. Intra-articular administration of gelatin hydrogels incorporating rapamycinemicelles reduces the development of experimental osteoarthritis in a murine model. *Biomaterials* **2014**, *35*, 9904–9911. [[CrossRef](#)] [[PubMed](#)]
133. Nims, R.J.; Cigan, A.D.; Albro, M.B.; Hung, C.T.; Ateshian, G.A. Synthesis rates and binding kinetics of matrix products in engineered cartilage constructs using chondrocyte-seeded agarose gels. *J. Biomech.* **2014**, *47*, 2165–2172. [[CrossRef](#)] [[PubMed](#)]
134. Sun, J.; Tan, H. Alginate-based biomaterials for regenerative medicine applications. *Materials* **2013**, *6*, 1285–1309. [[CrossRef](#)] [[PubMed](#)]
135. Shi, Y.; Xiong, D.S.; Peng, Y.; Wang, N. Effects of polymerization degree on recovery behavior of PVA/PVP hydrogels as potential articular cartilage prosthesis after fatigue test. *eXPRESS Polym. Lett.* **2016**, *10*, 125–138. [[CrossRef](#)]
136. Bichara, D.A.; Bodugoz-Sentruk, H.; Ling, D.; Malchau, E.; Bragdon, C.R.; Muratoglu, O.K. Osteochondral defect repair using a polyvinyl alcohol-polyacrylic acid (PVA-PAAc) hydrogel. *Biomed. Mater.* **2014**, *9*. [[CrossRef](#)] [[PubMed](#)]
137. Yang, H.Y.; van Ee, R.J.; Timmer, K.; Craenmehr, E.G.M.; Huang, J.H.; Öner, F.C.; Dhert, W.J.A.; Kragten, A.H.M.; Willems, N.; Grinwis, G.C.M.; et al. A novel injectable thermoresponsive and cytocompatible gel of poly(*N*-isopropylacrylamide) with layered double hydroxides facilitates siRNA delivery into chondrocytes in 3D culture. *Acta Biomater.* **2015**, *23*, 214–228. [[CrossRef](#)] [[PubMed](#)]
138. Yang, S.S.; Choi, W.H.; Song, B.R.; Jin, H.; Lee, S.J.; Lee, S.H.; Lee, J.; Kim, Y.J.; Park, S.R.; Park, S.-H.; et al. Fabrication of an osteochondral graft with using a solid freeform fabrication system. *Tissue Eng. Regen. Med.* **2015**, *12*, 239–248. [[CrossRef](#)]
139. Kim, J.E.; Kim, S.H.; Jung, Y. In situ chondrogenic differentiation of bone marrow stromal cells in bioactive self-assembled peptide gels. *J. Biosci. Bioeng.* **2015**, *120*, 91–98. [[CrossRef](#)] [[PubMed](#)]
140. Park, Y.B.; Song, M.; Lee, C.H.; Kim, J.A.; Ha, C.W. Cartilage repair by human umbilical cord blood-derived mesenchymal stem cells with different hydrogels in a rat model. *J. Orthop. Res.* **2015**, 1580–1586. [[CrossRef](#)] [[PubMed](#)]
141. Mao, H.; Kawazoe, N.; Chen, G. Cellular uptake of single-walled carbon nanotubes in 3D extracellular matrix-mimetic composite collagen hydrogels. *J. Nanosci. Nanotechnol.* **2014**, *14*, 2487–2492. [[CrossRef](#)] [[PubMed](#)]
142. Aberle, T.; Franke, K.; Rist, E.; Benz, K.; Schlosshauer, B. Cell-type specific four-component hydrogel. *PLoS ONE* **2014**, *9*, e86740. [[CrossRef](#)] [[PubMed](#)]
143. Chung, J.Y.; Song, M.; Ha, C.W.; Kim, J.A.; Lee, C.H.; Park, Y.B. Comparison of articular cartilage repair with different hydrogel-human umbilical cord blood-derived mesenchymal stem cell composites in a rat model. *Stem Cell Res. Ther.* **2014**, *5*, 39. [[CrossRef](#)] [[PubMed](#)]

144. Salamon, A.; van Vlierberghe, S.; van Nieuwenhove, I.; Baudisch, F.; Graulus, G.-J.; Benecke, V.; Alberti, K.; Neumann, H.-G.; Rychly, J.; Martins, J.C.; et al. Gelatin-based hydrogels promote chondrogenic differentiation of human adipose tissue-derived mesenchymal stem cells in vitro. *Materials* **2014**, *7*, 1342–1359. [[CrossRef](#)] [[PubMed](#)]
145. Fisher, M.B.; Henning, E.A.; Söegaard, N.B.; Dodge, G.R.; Steinberg, D.R.; Mauck, R.L. Maximizing cartilage formation and integration via a trajectory-based tissue engineering approach. *Biomaterials* **2014**, *35*, 2140–2148. [[CrossRef](#)] [[PubMed](#)]
146. Song, K.; Li, L.; Li, W.; Zhu, Y.; Jiao, Z.; Lim, M.; Fang, M.; Shi, F.; Wang, L.; Liu, T. Three-dimensional dynamic fabrication of engineered cartilage based on chitosan/gelatin hybrid hydrogel scaffold in a spinner flask with a special designed steel frame. *Mater. Sci. Eng. C* **2015**, *55*, 384–392. [[CrossRef](#)] [[PubMed](#)]
147. Mhanna, R.; Kashyap, A.; Palazzolo, G.; Vallmajo-Martin, Q.; Becher, J.; Möller, S.; Schnabelrauch, M.; Zenobi-Wong, M. Chondrocyte culture in three dimensional alginate sulfate hydrogels promotes proliferation while maintaining expression of chondrogenic markers. *Tissue Eng. A* **2014**, *20*, 1454–1464. [[CrossRef](#)] [[PubMed](#)]
148. Kesti, M.; Müller, M.; Becher, J.; Schnabelrauch, M.; D’Este, M.; Eglin, D.; Zenobi-Wong, M. A versatile bioink for three-dimensional printing of cellular scaffolds based on thermally and photo-triggered tandem gelation. *Acta Biomater.* **2015**, *11*, 162–172. [[CrossRef](#)] [[PubMed](#)]
149. Ishikawa, M.; Yoshioka, K.; Urano, K.; Tanaka, Y.; Hatanaka, T.; Nii, A. Biocompatibility of cross-linked hyaluronate (Gel-200) for the treatment of knee osteoarthritis. *Osteoarthr. Cartil.* **2014**, *22*, 1902–1909. [[CrossRef](#)] [[PubMed](#)]
150. Mazaki, T.; Shiozaki, Y.; Yamane, K.; Yoshida, A.; Nakamura, M.; Yoshida, Y.; Zhou, D.; Kitajima, T.; Tanaka, M.; Ito, Y.; et al. A novel, visible light-induced, rapidly cross-linkable gelatin scaffold for osteochondral tissue engineering. *Sci. Rep.* **2014**, *4*, 4457. [[CrossRef](#)] [[PubMed](#)]
151. Zhao, M.; Chen, Z.; Liu, K.; Wan, Y.Q.; Li, X.D.; Luo, X.W.; Bai, Y.G.; Yang, Z.L.; Feng, G. Repair of articular cartilage defects in rabbits through tissue-engineered cartilage constructed with chitosan hydrogel and chondrocytes. *J. Zhejiang Univ.-Sci. B (Biomed. Biotechnol.)* **2015**, *16*, 914–923. [[CrossRef](#)] [[PubMed](#)]
152. Kyomoto, M.; Moro, T.; Yamane, S.; Hashimoto, M.; Takatori, Y.; Ishihara, K. Effect of UV-irradiation intensity on graft polymerization of 2-methacryloyloxyethyl phosphorylcholine on orthopedic bearing substrate. *Mater. Res. Part A* **2014**, *102A*, 3012–3023. [[CrossRef](#)] [[PubMed](#)]
153. Sardinha, V.M.; Lima, L.L.; Belangero, W.D.; Zavaglia, C.A.; Bavaresco, V.P.; Gomes, J.R. Tribological characterization of polyvinyl alcohol hydrogel as substitute of articular cartilage. *Wear* **2013**, *301*, 218–225. [[CrossRef](#)]
154. Shi, Y.; Xiong, D. Microstructure and friction properties of PVA/PVP hydrogels for articular cartilage repair as function of polymerization degree and polymer concentration. *Wear* **2013**, *35*, 280–285. [[CrossRef](#)]
155. Chen, T.; Hilton, M.J.; Brown, E.B.; Zuscik, M.J.; Awad, H.A. Engineering superficial zone features in tissue engineered cartilage. *Biotechnol. Bioeng.* **2013**, *110*, 1476–1486. [[CrossRef](#)] [[PubMed](#)]
156. Blum, M.M.; Ovaert, T.C. Low friction hydrogel for articular cartilage repair: Evaluation of mechanical and tribological properties in comparison with natural cartilage tissue. *Mater. Sci. Eng. C* **2013**, *33*, 4377–4383. [[CrossRef](#)] [[PubMed](#)]
157. Fan, C.; Liao, L.; Zhang, C.; Liu, L. A tough double network hydrogel for cartilage tissue engineering. *J. Mater. Chem. B* **2013**, *1*, 4251–4258. [[CrossRef](#)]
158. Blum, M.M.; Ovaert, T.C. Investigation of friction and surface degradation of innovative boundary lubricant functionalized hydrogel material for use as artificial articular cartilage. *Wear* **2013**, *301*, 201–209. [[CrossRef](#)]
159. Giavaresi, G.; Bondioli, E.; Melandri, D.; Giardino, R.; Tschon, M.; Torricelli, P.; Cenacchi, G.; Rotini, R.; Castagna, A.; Veronesi, F.; et al. Response of human chondrocytes and mesenchymal stromal cells to a decellularized human dermis. *BMC Musculoskelet. Disord.* **2013**, *14*, 12. [[CrossRef](#)] [[PubMed](#)]
160. Stocco, E.; Barbon, S.; Dalzoppo, D.; Lora, S.; Sartore, L.; Folini, M.; Parnigotto, P.P.; Grandi, C. Tailored PVA/ECM scaffolds for cartilage regeneration. *Biomed. Res. Int.* **2014**, *2014*, 762189. [[CrossRef](#)] [[PubMed](#)]
161. Baek, J.H.; Kim, K.; Yang, S.S.; Park, S.H.; Song, B.R.; Yun, H.W.; Jeong, S.I.; Kim, Y.J.; Min, B.H.; Kim, M.S. Preparation of extracellular matrix developed using porcine articular cartilage and in vitro feasibility study of porcine articular cartilage as an anti-adhesive film. *Materials* **2016**, *9*, 49. [[CrossRef](#)] [[PubMed](#)]

162. Chen, X.; Zhang, F.; He, X.; Xu, Y.; Yang, Z.; Chen, L.; Zhou, S.; Yang, Y.; Zhou, Z.; Sheng, W.; et al. Chondrogenic differentiation of umbilical cord-derived mesenchymal stem cells in type I collagen-hydrogel for cartilage engineering. *Inj. Int. J. Care Inj.* **2013**, *44*, 540–549. [[CrossRef](#)] [[PubMed](#)]
163. Wu, L.; Gonzalez, S.; Shah, S.; Kyupelyan, L.; Petrigliano, F.A.; McAllister, D.R.; Adams, J.S.; Karperien, M.; Tuan, T.L.; Benya, P.D.; et al. Extracellular matrix domain formation as an indicator of chondrocyte dedifferentiation and hypertrophy. *Tissue Eng. Part C* **2014**, *20*, 160–168. [[CrossRef](#)] [[PubMed](#)]
164. Lam, J.; Lu, S.; Lee, E.J.; Trachtenberg, J.E.; Meretoja, V.V.; Dahlin, R.L.; van den Beucken, J.J.; Tabata, Y.; Wong, M.E.; Jansen, J.A.; et al. Osteochondral defect repair using bilayered hydrogels encapsulating both chondrogenically and osteogenically pre-differentiated mesenchymal stem cells in a rabbit model. *Osteoarthr. Cartil.* **2014**, *22*, 1291–1300. [[CrossRef](#)] [[PubMed](#)]
165. Lu, H.-T.; Sheu, M.-T.; Lin, Y.-F.; Lan, J.; Chin, Y.-P.; Hsieh, M.-S.; Cheng, C.-W.; Chen, C.-H. Injectable hyaluronic-acid-doxycycline hydrogel therapy in experimental rabbit osteoarthritis. *Vet. Res.* **2013**, *9*, 68. [[CrossRef](#)] [[PubMed](#)]
166. Matsiko, A.; Gleeson, J.P.; O'Brien, F.J. Scaffold mean pore size influences mesenchymal stem cell chondrogenic differentiation and matrix deposition. *Tissue Eng. Part A* **2015**, *21*, 486–497. [[CrossRef](#)] [[PubMed](#)]
167. Hui, J.H.; Ren, X.; Afizah, M.H.; Chian, K.S.; Mikos, A.G. Oligo[poly(ethylene glycol)fumarate] hydrogel enhances osteochondral repair in porcine femoral condyle defects. *Clin. Orthop. Relat. Res.* **2013**, *471*, 1174–1185. [[CrossRef](#)] [[PubMed](#)]
168. Kwon, H.; Rainbow, R.S.; Sun, L.; Hui, C.K.; Cairns, D.M.; Preda, R.C.; Kaplan, D.L.; Zeng, L. Scaffold structure and fabrication method affect proinflammatory milieu in three-dimensional-cultured chondrocytes. *J. Biomed. Mater. Res. Part A* **2015**, *103A*, 534–544. [[CrossRef](#)] [[PubMed](#)]
169. Steele, J.A.; McCullen, S.D.; Callanan, A.; Autefage, H.; Accardi, M.A.; Dini, D.; Stevens, M.M. Combinatorial scaffold morphologies for zonal articular cartilage engineering. *Acta Biomater.* **2014**, *10*, 2065–2075. [[CrossRef](#)] [[PubMed](#)]
170. Perrier-Groult, E.; Padeloup, M.; Malbouyres, M.; Galéra, P.; Mallein-Gerin, F. Control of collagen production in mouse chondrocytes by using a combination of bone morphogenetic protein-2 and small interfering RNA targeting Col1a1 for hydrogel-based tissue-engineered cartilage. *Tissue Eng. Part C* **2013**, *19*, 652–664. [[CrossRef](#)] [[PubMed](#)]
171. Sridhar, B.V.; Doyle, N.R.; Randolph, M.A.; Anseth, K.S. Covalently tethered TGF- β 1 with encapsulated chondrocytes in a PEG hydrogel system enhances extracellular matrix production. *J. Biomed. Mater. Res. Part A* **2014**, *102A*, 4464–4472. [[CrossRef](#)] [[PubMed](#)]
172. Simson, J.A.; Strehin, I.A.; Lu, Q.; Uy, M.O.; Elisseeff, J.H. An adhesive bone marrow scaffold and bone morphogenetic-2 protein carrier for cartilage tissue engineering. *Biomacromolecules* **2013**, *14*, 637–643. [[CrossRef](#)] [[PubMed](#)]
173. Ayerst, B.I.; Day, A.J.; Nurcombe, V.; Cool, S.M.; Merry, C.L. New strategies for cartilage regeneration exploiting selected glycosaminoglycans to enhance cell fate determination. *Biochem. Soc. Trans.* **2014**, *42*, 703–709. [[CrossRef](#)] [[PubMed](#)]
174. Ahearne, M.; Kelly, D.J. A comparison of fibrin, agarose and gellan gum hydrogels as carriers of stem cells and growth factor delivery microspheres for cartilage regeneration. *Biomed. Mater.* **2013**, *8*, 035004. [[CrossRef](#)] [[PubMed](#)]
175. Lee, W.D.; Hurtig, M.B.; Pilliar, R.M.; Stanford, W.L.; Kandel, R.A. Engineering of hyaline cartilage with a calcified zone using bone marrow stromal cells. *Osteoarthr. Cartil.* **2015**, *23*, 1307–1315. [[CrossRef](#)] [[PubMed](#)]
176. Kaderli, S.; Boulocher, C.; Pillet, E.; Watrelot-Virieux, D.; Rougemont, A.L.; Roger, T.; Viguier, E.; Gurny, R.; Scapozza, L.; Jordan, O. A novel biocompatible hyaluronic acid–chitosan hybrid hydrogel for osteoarthritis therapy. *Int. J. Pharm.* **2015**, *483*, 158–168. [[CrossRef](#)] [[PubMed](#)]
177. Medved, F.; Gonser, P.; Lotter, O.; Albrecht, D.; Amr, A.; Schaller, H.E. Severe posttraumatic radiocarpal cartilage damage: First report of autologous chondrocyte implantation. *Arch. Orthop. Trauma Surg.* **2013**, *133*, 1469–1475. [[CrossRef](#)] [[PubMed](#)]
178. Niemietz, T.; Zass, G.; Hagmann, S.; Diederichs, S.; Gotterbarm, T.; Richter, W. Xenogeneic transplantation of articular chondrocytes into full-thickness articular cartilage defects in minipigs: Fate of cells and the role of macrophages. *Cell Tissue Res.* **2014**, *358*, 749–761. [[CrossRef](#)] [[PubMed](#)]

179. Adachi, N.; Ochi, M.; Deie, M.; Nakamae, A.; Kamei, G.; Uchio, Y.; Iwasa, J. Implantation of tissue-engineered cartilage-like tissue for the treatment for full-thickness cartilage defects of the knee. *Knee Surg. Sports Traumatol. Arthrosc.* **2014**, *22*, 1241–1248. [[CrossRef](#)] [[PubMed](#)]
180. Buwalda, S.J.; Boere, K.W.; Dijkstra, P.J.; Feijen, J.; Vermonden, T.; Hennink, W.E. Hydrogels in a historical perspective: From simple networks to smart materials. *J. Control. Release* **2014**, *190*, 254–273. [[CrossRef](#)] [[PubMed](#)]
181. Vikingsson, L.; Gallego Ferrer, G.; Gómez-Tejedor, J.A.; Gómez Ribelles, J.L. An “in vitro” experimental model to predict the mechanical behavior of macroporous scaffolds implanted in articular cartilage. *J. Mech. Behav. Biomed. Mater.* **2014**, *32*, 125–131. [[CrossRef](#)] [[PubMed](#)]
182. Yusong, P.; Qianqian, S.; Chengling, P.; Jing, W. Prediction of mechanical properties of multilayer gradient hydroxyapatite reinforced poly(vinyl alcohol) gel biomaterial. *J. Biomed. Mater. Res. Part B* **2013**, *101B*, 729–735. [[CrossRef](#)] [[PubMed](#)]
183. Pan, Y.; Xiong, D. Stress-relaxation models of nano-HA/PVA gel biocomposites. *Mech. Time-Depend. Mater.* **2013**, *17*, 195–204. [[CrossRef](#)]
184. Du, G.; Gao, G.; Hou, R.; Cheng, Y.; Chen, T.; Fu, J. Tough and fatigue resistant biomimetic hydrogels of interlaced self-assembled conjugated polymer belts with a polyelectrolyte network. *Chem. Mater.* **2014**, *26*, 3522–3529. [[CrossRef](#)]
185. Cao, Yi.; Xiong, D.; Niu, Y.; Mei, Y.; Yin, Z.; Gui, J. Compressive properties and creep resistance of a novel, porous, semidegradable poly(vinyl alcohol)/poly(lactic-co-glycolic acid) scaffold for articular cartilage repair. *J. Appl. Polym. Sci.* **2014**, *131*, 40311. [[CrossRef](#)]
186. Gonzalez, J.S.; Alvarez, V.A. Mechanical properties of polyvinyl alcohol/hydroxyapatite cryogel as potential artificial cartilage. *J. Mech. Behav. Biomed. Mater.* **2014**, *34*, 47–56. [[CrossRef](#)] [[PubMed](#)]
187. Chen, K.; Zhang, D.; Dai, Z.; Wang, S.; Ge, S. Research on the interstitial fluid load support characteristics and start-up friction mechanisms of PVA-HA-silk composite hydrogel. *J. Bionic Eng.* **2014**, *11*, 378–388. [[CrossRef](#)]
188. Mansour, J.M.; Gu, D.W.; Chung, C.Y.; Heebner, J.; Althans, J.; Abdalian, S.; Schluchter, M.D.; Liu, Y.; Welter, J.F. Towards the feasibility of using ultrasound to determine mechanical properties of tissues in a bioreactor. *Ann. Biomed. Eng.* **2014**, *40*, 2190–2202. [[CrossRef](#)] [[PubMed](#)]
189. Yun, A.; Lee, S.-H.; Kim, J. A phase-field model for articular cartilage regeneration in degradable scaffolds. *Bull. Math. Biol.* **2013**, *75*, 2389–2409. [[CrossRef](#)] [[PubMed](#)]
190. Sandker, M.J.; Petit, A.; Redout, E.M.; Siebelt, M.; Müller, B.; Bruin, P.; Meyboom, R.; Vermonden, T.; Hennink, W.E.; Weinans, H. In situ forming acyl-capped PCLA-PEG-PCLA triblock copolymer based hydrogels. *Biomaterials* **2013**, *34*, 8002–8011. [[CrossRef](#)] [[PubMed](#)]
191. Fukui, T.; Kitamura, N.; Kurokawa, T.; Yokota, M.; Kondo, E.; Gong, J.P.; Yasuda, K. Intra-articular administration of hyaluronic acid increases the volume of the hyaline cartilage regenerated in a large osteochondral defect by implantation of a double-network gel. *J. Mater. Sci. Mater. Med.* **2014**, *25*, 1173–1182. [[CrossRef](#)] [[PubMed](#)]
192. Haaparanta, A.M.; Järvinen, E.; Cengiz, I.F.; Ellä, V.; Kokkonen, H.T.; Kiviranta, I.; Kellomäki, M. Preparation and characterization of collagen/PLA, chitosan/PLA, and collagen/chitosan/PLA hybrid scaffolds for cartilage tissue engineering. *J. Mater. Sci. Mater. Med.* **2014**, *25*, 1129–1136. [[CrossRef](#)] [[PubMed](#)]
193. Lin, H.; Cheng, A.W.; Alexander, P.G.; Beck, A.M.; Tuan, R.S. Cartilage tissue engineering application of injectable gelatin hydrogel with in situ visible-light-activated gelation capability in both air and aqueous solution. *Tissue Eng. Part A* **2014**, *2014*, 2402–2411. [[CrossRef](#)] [[PubMed](#)]
194. Li, X.; Li, Y.; Zuo, Y.; Qu, D.; Liu, Y.; Chen, T.; Jiang, N.; Li, H.; Li, J. Osteogenesis and chondrogenesis of biomimetic integrated porous PVA/gel/V-n-HA/pa6 scaffolds and BMSCs construct in repair of articular osteochondral defect. *J. Biomed. Mater. Res. Part A* **2015**, *103A*, 3226–3236. [[CrossRef](#)] [[PubMed](#)]
195. Hoch, E.; Tovar, G.E.; Borchers, K. Biopolymer-based hydrogels for cartilage tissue engineering. *Bioinspir. Biomim. Nanobiomater.* **2016**, *5*, 51–66. [[CrossRef](#)]
196. Tuan, R.S.; Chen, A.F.; Klatt, B.A. Cartilage regeneration. *J. Am. Acad. Orthop. Surg.* **2013**, *21*, 303–311. [[CrossRef](#)] [[PubMed](#)]
197. Sivashanmugam, A.; Kumar, R.A.; Priya, M.V.; Nair, S.V.; Jayakumar, R. An overview of injectable polymeric hydrogels for tissue engineering. *Eur. Polym. J.* **2015**, *72*, 543–565. [[CrossRef](#)]

198. Hu, X.; Li, D.; Zhou, F.; Gao, C. Biological hydrogel synthesized from hyaluronic acid, gelatin and chondroitin sulfate by click chemistry. *Acta Biomater.* **2011**, *7*, 1618–1626. [[CrossRef](#)] [[PubMed](#)]
199. Yang, J.-A.; Yeom, J.; Hwang, B.W.; Hoffman, A.S.; Hahn, S.K. In situ-forming injectable hydrogels for regenerative medicine. *Prog. Polym. Sci.* **2014**, *39*, 1973–1986. [[CrossRef](#)]
200. Mirahmadi, F.; Tafazzoli-Shadpour, M.; Shokrgozar, M.A.; Bonakdar, S. Enhanced mechanical properties of thermosensitive chitosan hydrogel by silk fibers for cartilage tissue engineering. *Mater. Sci. Eng. C* **2013**, *33*, 4786–4794. [[CrossRef](#)] [[PubMed](#)]
201. Moreira Teixeira, L.S.; Patterson, J.; Luyten, F.P. Skeletal tissue regeneration: Where can hydrogels play a role? *Int. Orthop.* **2014**, *38*, 1861–1876. [[CrossRef](#)] [[PubMed](#)]
202. Camci-Unal, G.; Cuttica, D.; Annabi, N.; Demarchi, D.; Khademhosseini, A. Synthesis and characterization of hybrid hyaluronic acid-gelatin hydrogels. *Biomacromolecules* **2013**, *14*, 1085–1092. [[CrossRef](#)] [[PubMed](#)]
203. Pot, M.W.; Faraj, K.A.; Adawy, A.; van Enkevort, W.J.; van Moerkerk, H.T.; Vlieg, E.; Daamen, W.F.; van Kuppevelt, T.H. Versatile wedge-based system for the construction of unidirectional collagen scaffolds by directional freezing: Practical and theoretical considerations. *Appl. Mater. Interfaces* **2015**, *7*, 8495–8505. [[CrossRef](#)] [[PubMed](#)]
204. Mao, H.; Kawazoe, N.; Chen, G. Cell response to single-walled carbon nanotubes in hybrid porous collagen sponges. *Colloids Surf. B Biointerfaces* **2015**, *126*, 63–69. [[CrossRef](#)] [[PubMed](#)]
205. Daly, A.C.; Critchley, S.E.; Rencsok, E.M.; Kelly, D.J. A comparison of different bioinks for 3D bioprinting of fibrocartilage and hyaline cartilage. *Biofabrication* **2016**, *8*, 045002. [[CrossRef](#)] [[PubMed](#)]
206. Seol, Y.J.; Park, J.Y.; Jeong, W.; Kim, T.H.; Kim, S.Y.; Cho, D.W. Development of hybrid scaffolds using ceramic and hydrogel for articular cartilage tissue regeneration. *J. Biomed. Mater. Res. Part A* **2015**, *103A*, 1404–1413. [[CrossRef](#)] [[PubMed](#)]
207. Xu, T.; Binder, K.W.; Albanna, M.Z.; Dice, D.; Zhao, W.; Yoo, J.J.; Atala, A. Hybrid printing of mechanically and biologically improved constructs for cartilage tissue engineering applications. *Biofabrication* **2013**, *5*, 015001. [[CrossRef](#)] [[PubMed](#)]
208. Vázquez-Portalatín, N.; Kilmer, C.E.; Panitch, A.; Liu, J.C. Characterization of collagen type I and II blended hydrogels for articular cartilage tissue engineering. *Biomacromolecules* **2016**, *17*, 3145–3152. [[CrossRef](#)] [[PubMed](#)]
209. Mintz, B.R.; Cooper, J.A., Jr. Hybrid hyaluronic acid hydrogel/poly(ϵ -caprolactone) scaffold provides mechanically favorable platform for cartilage tissue engineering studies. *J. Biomed. Mater. Res. Part A* **2014**, *102A*, 2918–2926. [[CrossRef](#)] [[PubMed](#)]
210. Sun, W.; Xue, B.; Li, Y.; Qin, M.; Wu, J.; Lu, K.; Wu, J.; Cao, Y.; Jiang, Q.; Wang, W. Polymer-supramolecular polymer double-network hydrogel. *Adv. Funct. Mater.* **2016**, *26*, 9044–9052. [[CrossRef](#)]
211. Formica, F.A.; Öztürk, E.; Hess, S.C.; Stark, W.J.; Maniura-Weber, K.; Rottmar, M.; Zenobi-Wong, M. A bioinspired ultraporos nanofiber-hydrogel mimic of the cartilage extracellular matrix. *Adv. Funct. Mater.* **2016**, *5*, 3129–3138. [[CrossRef](#)] [[PubMed](#)]
212. Kang, M.L.; Jeong, S.Y.; Im, G.I. Hyaluronic acid hydrogel functionalized with self-assembled micelles of amphiphilic PEGylated kartogenin for the treatment of osteoarthritis. *Tissue Eng. Part A* **2017**, *23*, 630–639. [[CrossRef](#)] [[PubMed](#)]
213. Huang, H.; Zhang, X.; Hu, X.; Dai, L.; Zhu, J.; Man, Z.; Chen, H.; Zhou, C.; Ao, Y. Directing chondrogenic differentiation of mesenchymal stem cells with a solid-supported chitosan thermogel for cartilage tissue engineering. *Biomed. Mater.* **2014**, *9*, 035008. [[CrossRef](#)] [[PubMed](#)]
214. Buchtová, N.; Réthoré, G.; Boyer, C.; Guicheux, J.; Rambaud, F.; Vallé, K.; Belleville, P.; Sanchez, C.; Chauvet, O.; Weiss, P.; et al. Nanocomposite hydrogels for cartilage tissue engineering: Mesoporous silica nanofibers interlinked with siloxane derived polysaccharide. *J. Mater. Sci. Mater. Med.* **2013**, *24*, 1875–1884. [[CrossRef](#)] [[PubMed](#)]
215. Wang, T.; Lai, J.H.; Yang, F. Effects of hydrogel stiffness and extracellular compositions on modulating regeneration by mixed populations of stem cells and chondrocytes in vivo. *Tissue Eng. Part A* **2016**, *22*, 1348–1356. [[CrossRef](#)] [[PubMed](#)]
216. Gyles, D.A.; Castro, L.D.; Silva, J.O.C., Jr.; Ribiero-Costa, R.M. A review of the designs and prominent biomedical advances of natural and synthetic hydrogel formulations. *Eur. Polym. J.* **2017**, *88*, 373–392. [[CrossRef](#)]

217. Jeon, S.-J.; Hauser, A.W.; Hayward, R.C. Shape-morphing materials from stimuli-responsive hydrogel hybrids. *Acc. Chem. Res.* **2017**, *50*, 161–169. [[CrossRef](#)] [[PubMed](#)]
218. Jeon, J.E.; Vaquette, C.; Theodoropoulos, C.; Klein, T.J.; Hutmacher, D.W. Multiphasic construct studied in an ectopic osteochondral defect model. *J. R. Soc. Interface* **2014**, *11*, 20140184. [[CrossRef](#)] [[PubMed](#)]
219. Childs, A.; Hemraz, U.D.; Castro, N.J.; Fenniri, H.; Zhang, L.G. Novel biologically-inspired rosette nanotube PLLA scaffolds for improving human mesenchymal stem cell chondrogenic differentiation. *Biomed. Mater.* **2013**, *8*, 065003. [[CrossRef](#)] [[PubMed](#)]
220. Mačiulaitis, J.; Deveikytė, M.; Rekšytė, S.; Bratchikov, M.; Darinskas, A.; Šimbelytė, A.; Daunoras, G.; Laurinavičienė, A.; Laurinavičius, A.; Gudas, R.; et al. Preclinical study of SZ20180 material 3D microstructured scaffolds for cartilage tissue engineering made by femtosecond direct laser writing lithography. *Biofabrication* **2015**, *7*, 015015. [[CrossRef](#)] [[PubMed](#)]
221. Heinemann, S.; Coradin, T.; Desimone, M.F. Bio-inspired silica-collagen materials: Applications and perspectives in the medical field. *Biomater. Sci.* **2013**, *1*, 688–702. [[CrossRef](#)]



© 2017 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).