

Review Article

Stem cells as cellular vehicles for gene therapy against glioblastoma

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Received August 7, 2015; Accepted October 5, 2015; Epub October 15, 2015; Published October 30, 2015

Abstract: Glioblastoma (GBM) is the most common and deadliest primary tumor in adults, with current treatments having limited specific and efficient delivery of therapeutic drugs to tumor sites or cells. Therefore, the development of alternative treatment options is urgently needed. Stem cells are considered as ideal cellular vehicles for gene therapy against glioblastoma. In this paper, we reviewed the recent studies investigating the use of different types of stem cells as cellular vehicles and the gene of interests against the glioblastoma, as well as the future directions of the application of cellular vehicles mediated therapy for glioblastoma.

Keywords: Glioma, stem cell, cellular vehicle, gene therapy

Introduction

Glioblastoma makes 25% of all malignant nervous system tumor, which occurs about 3 per 100,000 population in United States [1]. Because of its aggressive characteristics and low specific and efficient delivery of therapeutic drugs to tumor sites, treatments for glioblastoma include surgery, radiotherapy and chemotherapy lead to only 25% of patients surviving about 2 years even with advanced technology [2]. Thus, the development of better therapeutic strategies to enhance the survival rate is desperately needed [3].

Stem cells are a group of cells with self-renewal and multilineage differentiation. A large number of studies have demonstrated that stem cells derived from various sources could specifically migrate to tumor sites [4-7]. Therefore, stem cell is a promising vehicle loaded with anti-tumor drugs or gene of interests against tumor home to tumor site [8].

Stem cells as cellular vehicles

Neural stem cells

Neural stem cells (NSCs) are central neural system (CNS) progenitor cells, which have self-

renewal ability and can differentiate into all of the three types of cells in CNS: neurons, oligodendrocytes and astrocytes [9]. Recent studies showed that NSCs could migrate through the brain and target to tumor site, which indicated that NSCs was a cellular vehicle for delivering anti-brain therapeutic drugs or gene of interests [10]. The possible mechanism was that cytokines and other factors produced in the tumor microenvironment, such as hepatocyte growth factor (HGF), hypoxia-inducible factor-1 α (HIF-1 α) and vascular endothelial growth factor (VEGF) act as chemoattractants for NSCs [11]. On the other hand, several studies have demonstrated that injected NSCs release soluble molecules to promote immune modulation in the CNS [12, 13]. Because NSCs reduce the activity of immune system, they are easy to carry and deliver anti-tumor drug and gene of interests.

Mesenchymal stem cells

Mesenchymal stem cells (MSCs) are multipotent cells which is able to differentiate into a number of cell types. These cells have been another potential vehicle for delivering anti-tumor therapy [14]. MSCs can be expanded *in*

in vitro with a low intrinsic mutation rate, are manipulated easily and well tolerant to the human immune system. They can also be autologously transplanted into the same patient where therapy taken to avoid immune response after administration [15]. Interestingly, MSCs have been proved to migrate into many different types of tumor microenvironments including glioma [16-18]. However, the exact migratory mechanism of MSCs is not completely understood. It is suspected that chemokines, cytokine, growth factors and metalloproteinase secreted by the tumor guide MSCs' tumor-targeting ability [19, 20]. Additionally, MSCs can be engineered to express homing ligands, which improves their target specificity [21].

Actually, bone marrow, adipose tissues, peripheral blood and embryonic stem cell-derived mesenchymal stem cells are the most common types [15, 22]. However, it has come up a new mesenchymal stem cells obtained from menstrual blood called menstrual blood-derived mesenchymal stem cells recently [23, 24]. These cells express stem cell markers such as SSEA-4, Oct-4, c-kit (CD117), and Nanog, and have the potent ability to differentiate into a variety of cell types, such as the heart, nerve, bone and liver [25, 26]. These cells secrete many growth factors to display recurrent angiogenesis [26]. This provides an easy way to get MSCs, indicating a potential application in cell carrier of gene therapy against glioma.

Gene of interest

Gene therapy for GBM is rapidly developing. The functions of genes of interest can result not only in tumor cell death, but also enhance immune responses to tumor antigens, as well as disruption of the tumor microenvironment, including inhibition of angiogenesis and neovascularization [27-29]. The genes of interest include the cytotoxic gene and immune stimulatory gene as follows.

Cytotoxic gene

Cytotoxic, radio- and chemotherapy have been the standard care for GBM patients. Most of the failure was because of their negative impacts on neighboring healthy tissue and small therapeutic indexes. Suicide gene therapy includes delivery of a prodrug activating enzyme (suicide gene) that is able to convert nontoxic prodrugs to cytotoxic forms [30].

Herpes simplex virus-thymidine kinase (HSV-TK)

HSV-TK/ganciclovir (GCV) is one of the most widely used prodrug activation systems. In this suicide gene therapy system, GCV is non-toxic and can readily cross the blood-brain barrier, converted into active drug in the tumor cells. Then GCV will be phosphorylated and incorporated into replicating DNA, leading to cell death. The phosphorylated GCV can pass through the gap junction of adjacent cells, and kill neighboring tumor cells. This ability was called the "bystander effect", which was defined as death of tumor cells adjacent to modified cells [31, 32]. As the bystander effect could cover the low transduction rate of retroviral system, clinical studies have used the retrovirus-mediated HSV-TK/GCV gene therapy, resulting in only clinical safety but not therapeutic benefits [33, 34].

However, in a previous study, a potent bystander effect between NSCs transduced with HSV-TK gene (NSCtk cells) was observed in intracranial tumor. NSCtk cells were injected at the intracranial site distant from the tumor implantation inducing the contralateral hemisphere to the tumor in rates. Results demonstrated a potent migratory and tumor hunting ability of NSCs and a potent *in vivo* anti-tumor effect of thymidine kinase through the bystander effect [31]. Furthermore, they found out that the strategy using mesenchymal stem cells transduced with HSV-TK (MSCtk cells) and ganciclovir (MSCtk therapy) is more feasible and practical for clinical application than the method using neural stem cells [35]. Recently, human embryonic stem cell-derived MSCs were evaluated as another alternative option in stem cells HSV-TK therapy [36].

Secretable trimeric form of tumor necrosis factor-related apoptosis-inducing ligand (stTRAIL)

Tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) based therapy strategy involves treatment with recombinant TRAIL (rTRAIL) or an adenovirus bearing the *TRAIL* gene anti glioma. The artificial *TRAIL* gene, that is secretable trimetric TRAIL (stTRAIL), encodes a fusion protein composed of three functional elements including a secretion signal, a trimerization domain and an apoptosis-inducing moiety of the *TRAIL* gene sequence [37]. Adenoviral vectors delivering the *stTRAIL* gene (Ad-stTRAIL)

had higher tumor suppressor rate compared with adenoviral vectors delivering the full-length sequence of *TRAIL* gene in vivo and in vitro [38]. Other studies also showed that TRAIL and secreted TRAIL dominant with or without fused to hFlt3L instead of stTRAIL also had profound anti-tumor effects in vivo [39-41].

Cytosine deaminase (CD)

Cytosine deaminase: uracil phosphoribosyl-transferase (CDy:UPRT) engineered human adipose tissue-derived mesenchymal stem cells (AT-MSCs), short as CDy-AT-MSCs, are able to convert non-toxic 5-fluorocytosine (5-FC) to the active toxic form [42]. Then they demonstrated that CDy-AT-MSCs/5-FC system of suicide gene therapy significantly inhibited glioblastoma growth [43, 44].

Interleukin-24 (IL-24)

The mda-7 gene, renamed as interleukin-24 (IL-24), was isolated from human melanoma cells induced to undergo terminal differentiation treating with fibroblast interferon and mezerein [45], which is a member of the interleukin-10 (IL-10) gene family [46-48]. Considering its cancer cells' specific apoptosis-inducing and tumor growth suppressing ability in human tumor animal models, mda-7/IL-24 was regarded in application in patients with advanced cancers [49, 50]. The pathways by which Ad.5-mda-7 causes cell death in tumor cells are not well understood, however, it seems that proteins important in the onset of growth inhibition and apoptosis, such as BCL-XL, BCL-2, and BAX [48, 51-53], are involved, which lead to mitochondrial dysfunction [54] and endoplasmic reticulum stress signaling [55].

A recent study employed a recombinant adenovirus that comprises the tail and shaft domains of a serotype 5 virus and the knob domain of a serotype 3 virus expressing MDA-7/IL-24 (Ad.5/3-mda-7) to combine both inhibition of cytoprotective pathways and tropism modification, and provided a means of developing an improved therapy for GBM [56].

Interleukin-13 (IL-13)

Human IL-13 was fused to the *Pseudomonas* exotoxin (hIL-13-PE; Cintredekin Besudotox) to target IL13R α 2-expressing GBM cells [57] as 50% to 80% of human GBMs express a number

of the IL-13 receptor, IL13R α 2 [58, 59]. A recent study developed Ad.mhIL-13-PE to provide sustained expression, effective anti-GBM cytotoxicity, and minimal neurotoxicity leading to a significant advance in the implementation of targeted toxins for glioma therapeutics [60].

EphrinA1-PE38

EphrinA1-PE38 is a specific immunotoxin against the EphA2 receptor which is a member of the Eph receptor tyrosine kinase family, whose 16 members can be further divided into "A" and "B" classes, based on sequence homology and binding affinity to their ligand, the Ephrin [61]. A recent study demonstrated that the intratumoral injection of hMSCs engineered with EphrinA1-PE38 was effective in inhibiting tumor growth in a glioma tumor model [62].

Immune stimulatory gene

The immune-privileged state of the brain is an important obstacle to immunotherapy against glioma [63]. The brain lacks antigen-presenting cells and is limited in lymphatics that impede immune cells from the brain parenchyma [64]. In addition, the GBM microenvironment is immunosuppressed, with elevated myeloid-derived suppressor cells and regulatory T cells [65]. Despite these challenges, significant progress with immunemediated gene therapy strategies has been achieved.

Interleukin-12 (IL-12)

IL-12 is one of the anti-tumor cytokines, driving from a T_H1 response [66]. A 34.5-deleted HSV-1 expressing mouse IL-12 (M002) was tested in non-human primates and proved to be nontoxic, but increased activation of nonhuman primates lymphocytes [67]. MSCs expressing IL-12 (MSC-IL12M) inhibited intracranial tumor growth and prolonged survival administered in the contralateral brain hemisphere [68].

Colony stimulating factor (CSF)

One of the immunotherapy strategies is to express cytokines to enhance adaptive immune system. JX-594 was a TK-deleted VV expressing granulocyte macrophage colony stimulating factor (GM-CSF) [69]. In two GBM models, JX-594 inhibited tumor growth and increased survival. It indicated to be linked with increased CSF-dependent inflammation [70].

Stem cell carriers for glioblastoma gene therapy

Fms-like tyrosine kinase 3 ligand (Flt3L)

Flt3L is a cytokine associated with the development of hematopoietic precursors into both conventional (cDCs) dendritic cells and plasmacytoid (pDCs), as well as their migration out of bone marrow [71]. Replication-defective Ad expressing Flt3L enhanced survival in a rat glioma model and this was linked with increased infiltration of DCs [72]. A similar strategy has been performed using oHSV expressing Flt3L. G47D-Flt3L significantly prolonged survival in the mouse glioma model [73].

Future directions

Considering the cell carriers' tropism to tumor cells, integrins and chemokines have been regarded as important for cell hunting for tumors sites [74]. Chemokines, which induce migration and manipulating integrin function to stimulate lymphocyte movement are used to attract various cell types to the tumor [75]. It has demonstrated that these chemokine systems can improve the efficacy of migrating immunotherapeutic cell carriers to tumors, and would be applied to improve the delivery of cell carriers to brain tumors [76].

Additionally, signaling pathways including uPA/uPAR (urokinase receptor), c-MET receptors and VEGF/VEGFR2 seem to play roles in the migration of stem cell to cancer cells [77]. The upregulation of these signaling pathways would improve the migratory activity of these cell carriers in glioma treatment.

Keeping the cell carriers alive around the tumor site for a longer period of time is another important strategy that will improve cell-carried therapies. Coating the carrier cells with a synthetic extracellular matrix (sECM) promotes stem cell survival. This approach reduced tumor volume when used with TRAIL engineered stem cells in vivo successfully [78]. These results are particularly attractive because the efficacy of the sECM surrounded NSCs was determined in a resected tumor cavity. This study also demonstrated that encapsulating MSCs with a biodegradable sECM enabled their retention in the tumor resection cavity and allowed them to release tumor suppressing therapy for a longer period of time.

At last, when cell carriers are loaded with a therapeutic virus, the virus would better to

remain quiescent until delivered to the tumor cells. Otherwise, the virus will destroy the cell carrier and potential therapeutic effect will be lost. Therefore, creating viruses that only replicate when they reach the tumor site will improve their clinical efficacy in patients with brain tumors.

Alternatively, differentiated cells can be induced into stem cell state. Cells derived from urine, and skin other tissues have been reprogrammed into induced pluripotent NSCs successfully [79]. These induced pluripotent stem (iPS) cell-derived NSCs have delivered gene therapy following contralateral intracranial injection in mice with glioma xenografts successfully [80]. Advantages of iPS cells instead of stem cells include their ability to escape from immune rejection and the absence of ethical concerns when using human embryonic cells. Additionally, iPS cells can be easily generated from somatic cells, which makes it excellent option for investigations in many model systems. However, these cells still have the potential for tumorigenicity. Furthermore, which somatic cells provide the best source to generate iPS cells or which reprogramming technique is the most efficient and safest still remains uncertain [81]. While the treatment of brain cancer is in infancy using iPS cells as carriers of therapeutic agents, the clinical potential of iPS cells is promising.

Conclusions

Even with the advanced therapies for glioma, patients are still faced with a poor prognosis. The low efficiency of delivering molecular therapies to the tumor site has in large part resulted in unremarkable benefits to glioma patients in clinical trials. Stem cells as cellular vehicles have been used to improve the delivery of these therapies. The natural tumor tropism, loading capability and modifiable characteristics of stem cells are major advantages that augment molecular therapies. The genes of interest include Cytotoxic gene and Immune stimulatory gene. With a better understanding of the potential tumor tropism mechanism, modifications can be performed to improve clinical efficacy of cell carrier system. To avoid ethical concerns and get stem cells relative easily, using induced pluripotent stem (iPS) cell is another direction in the future.

Acknowledgements

This work was supported by the National Science Foundation of China (Nos. 81201783 81372463, 81371954 and 81171687), Zhejiang Provincial Natural Science Foundation of China (Nos. LY15H160051, LY14H160041 and LY13H080005), Funds of Science Technology Department of Zhejiang Province (No. 2014-C37101), Zhejiang Province Bureau of Health (Nos. 2015ZA009 and WKJ-ZJ-1502), Funds of Science Technology Department of Zhejiang Province (No. 2014C37101), the National High-tech R&D Program (863 program, No. 2011AA020102 and No. 2012AA020905), the Key Technologies R&D Program of Zhejiang Province (No. 2012C03SA170003), and the Hangzhou Key Technologies R&D Program (No. 20122513A49).

Disclosure of conflict of interest

None.

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References

- [1] Brandes AA, Tosoni A, Franceschi E, Reni M, Gatta G, Vecht C. Glioblastoma in adults. *Crit Rev Oncol Hematol* 2008; 67: 139-152.
- [2] Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, Belanger K, Brandes AA, Marosi C, Bogdahn U, Curschmann J, Janzer RC, Ludwin SK, Gorlia T, Allgeier A, Lacombe D, Cairncross JG, Eisenhauer E, Mirimanoff RO. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N Engl J Med* 2005; 352: 987-996.
- [3] Westphal M, Lamszus K. The neurobiology of gliomas: from cell biology to the development of therapeutic approaches. *Nat Rev Neurosci* 2011; 12: 495-508.
- [4] Nakamizo A, Marini F, Amano T, Khan A, Studeny M, Gumin J, Chen J, Hentschel S, Vecil G, Dembinski J, Andreeff M, Lang FF. Human bone marrow-derived mesenchymal stem cells in the treatment of gliomas. *Cancer Res* 2005; 65: 3307-3318.
- [5] Balber AE. Concise review: aldehyde dehydrogenase bright stem and progenitor cell populations from normal tissues: characteristics, activities, and emerging uses in regenerative medicine. *Stem Cells* 2011; 29: 570-575.
- [6] Wu J, Li J, Zhang N, Zhang C. Stem cell-based therapies in ischemic heart diseases: a focus on aspects of microcirculation and inflammation. *Basic Res Cardiol* 2011; 106: 317-324.
- [7] Jiang Y, Jahagirdar BN, Reinhardt RL, Schwartz RE, Keene CD, Ortiz-Gonzalez XR, Reyes M, Lenvik T, Lund T, Blackstad M, Du J, Aldrich S, Lisberg A, Low WC, Largaespada DA, Verfaillie CM. Pluripotency of mesenchymal stem cells derived from adult marrow. *Nature* 2002; 418: 41-49.
- [8] Tabatabai G, Wick W, Weller M. Stem cell-mediated gene therapies for malignant gliomas: a promising targeted therapeutic approach? *Discov Med* 2011; 11: 529-536.
- [9] Temple S. The development of neural stem cells. *Nature* 2001; 414: 112-117.
- [10] Muller FJ, Snyder EY, Loring JF. Gene therapy: can neural stem cells deliver? *Nat Rev Neurosci* 2006; 7: 75-84.
- [11] An JH, Lee SY, Jeon JY, Cho KG, Kim SU, Lee MA. Identification of gliotropic factors that induce human stem cell migration to malignant tumor. *J Proteome Res* 2009; 8: 2873-2881.
- [12] Pluchino S, Quattrini A, Brambilla E, Gritti A, Salani G, Dina G, Galli R, Del CU, Amadio S, Bergami A, Furlan R, Comi G, Vescovi AL, Martino G. Injection of adult neurospheres induces recovery in a chronic model of multiple sclerosis. *Nature* 2003; 422: 688-694.
- [13] Pluchino S, Gritti A, Blezer E, Amadio S, Brambilla E, Borsellino G, Cossetti C, Del CU, Comi G, T HB, Vescovi A, Martino G. Human neural stem cells ameliorate autoimmune encephalomyelitis in non-human primates. *Ann Neurol* 2009; 66: 343-354.
- [14] Shah K. Mesenchymal stem cells engineered for cancer therapy. *Adv Drug Deliv Rev* 2012; 64: 739-748.
- [15] Jones BJ, McTaggart SJ. Immunosuppression by mesenchymal stromal cells: from culture to clinic. *Exp Hematol* 2008; 36: 733-741.
- [16] Nakamizo A, Marini F, Amano T, Khan A, Studeny M, Gumin J, Chen J, Hentschel S, Vecil G, Dembinski J, Andreeff M, Lang FF. Human bone marrow-derived mesenchymal stem cells in the treatment of gliomas. *Cancer Res* 2005; 65: 3307-3318.
- [17] Kim SM, Lim JY, Park SI, Jeong CH, Oh JH, Jeong M, Oh W, Park SH, Sung YC, Jeun SS. Gene therapy using TRAIL-secreting human umbilical cord blood-derived mesenchymal stem cells against intracranial glioma. *Cancer Res* 2008; 68: 9614-9623.
- [18] Hu YL, Huang B, Zhang TY, Miao PH, Tang GP, Tabata Y, Gao JQ. Mesenchymal stem cells as a novel carrier for targeted delivery of gene in cancer therapy based on nonviral transfection. *Mol Pharm* 2012; 9: 2698-2709.

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- [19] Beckermann BM, Kallifatidis G, Groth A, Frommhold D, Apel A, Mattern J, Salnikov AV, Moldenhauer G, Wagner W, Diehlmann A, Saffrich R, Schubert M, Ho AD, Giese N, Buchler MW, Friess H, Buchler P, Herr I. VEGF expression by mesenchymal stem cells contributes to angiogenesis in pancreatic carcinoma. *Br J Cancer* 2008; 99: 622-631.
- [20] Chamberlain G, Fox J, Ashton B, Middleton J. Concise review: mesenchymal stem cells: their phenotype, differentiation capacity, immunological features, and potential for homing. *Stem Cells* 2007; 25: 2739-2749.
- [21] Huang J, Zhang Z, Guo J, Ni A, Deb A, Zhang L, Mirotsov M, Pratt RE, Dzau VJ. Genetic modification of mesenchymal stem cells overexpressing CCR1 increases cell viability, migration, engraftment, and capillary density in the injured myocardium. *Circ Res* 2010; 106: 1753-1762.
- [22] Bak XY, Lam DH, Yang J, Ye K, Wei EL, Lim SK, Wang S. Human embryonic stem cell-derived mesenchymal stem cells as cellular delivery vehicles for prodrug gene therapy of glioblastoma. *Hum Gene Ther* 2011; 22: 1365-1377.
- [23] Patel AN, Park E, Kuzman M, Benetti F, Silva FJ, Allickson JG. Multipotent menstrual blood stromal stem cells: isolation, characterization, and differentiation. *Cell Transplant* 2008; 17: 303-311.
- [24] Patel AN, Silva F. Menstrual blood stromal cells: the potential for regenerative medicine. *Regen Med* 2008; 3: 443-444.
- [25] Mou XZ, Lin J, Chen JY, Li YF, Wu XX, Xiang BY, Li CY, Ma JM, Xiang C. Menstrual blood-derived mesenchymal stem cells differentiate into functional hepatocyte-like cells. *J Zhejiang Univ Sci B* 2013; 14: 961-972.
- [26] Lin J, Xiang D, Zhang JL, Allickson J, Xiang C. Plasticity of human menstrual blood stem cells derived from the endometrium. *J Zhejiang Univ Sci B* 2011; 12: 372-380.
- [27] Castro MG, Candolfi M, Kroeger K, King GD, Curtin JF, Yagiz K, Mineharu Y, Assi H, Wibowo M, Ghulam MA, Foulad D, Puntel M, Lowenstein PR. Gene therapy and targeted toxins for glioma. *Curr Gene Ther* 2011; 11: 155-180.
- [28] Assi H, Candolfi M, Baker G, Mineharu Y, Lowenstein PR, Castro MG. Gene therapy for brain tumors: basic developments and clinical implementation. *Neurosci Lett* 2012; 527: 71-77.
- [29] Samaranyake H, Maatta AM, Pikkarainen J, Yla-Herttuala S. Future prospects and challenges of antiangiogenic cancer gene therapy. *Hum Gene Ther* 2010; 21: 381-396.
- [30] Duarte S, Carle G, Faneca H, de Lima MC, Pierrefite-Carle V. Suicide gene therapy in cancer: where do we stand now? *Cancer Lett* 2012; 324: 160-170.
- [31] Li S, Gao Y, Tokuyama T, Yamamoto J, Yokota N, Yamamoto S, Terakawa S, Kitagawa M, Namba H. Genetically engineered neural stem cells migrate and suppress glioma cell growth at distant intracranial sites. *Cancer Lett* 2007; 251: 220-227.
- [32] Dilber MS, Abedi MR, Christensson B, Bjorkstrand B, Kidder GM, Naus CC, Gahrton G, Smith CI. Gap junctions promote the bystander effect of herpes simplex virus thymidine kinase in vivo. *Cancer Res* 1997; 57: 1523-1528.
- [33] Rainov NG. A phase III clinical evaluation of herpes simplex virus type 1 thymidine kinase and ganciclovir gene therapy as an adjuvant to surgical resection and radiation in adults with previously untreated glioblastoma multiforme. *Hum Gene Ther* 2000; 11: 2389-2401.
- [34] Ram Z, Culver KW, Oshiro EM, Viola JJ, DeVroom HL, Otto E, Long Z, Chiang Y, McGarrity GJ, Muul LM, Katz D, Blaese RM, Oldfield EH. Therapy of malignant brain tumors by intratumoral implantation of retroviral vector-producing cells. *Nat Med* 1997; 3: 1354-1361.
- [35] Amano S, Li S, Gu C, Gao Y, Koizumi S, Yamamoto S, Terakawa S, Namba H. Use of genetically engineered bone marrow-derived mesenchymal stem cells for glioma gene therapy. *Int J Oncol* 2009; 35: 1265-1270.
- [36] Bak XY, Lam DH, Yang J, Ye K, Wei EL, Lim SK, Wang S. Human embryonic stem cell-derived mesenchymal stem cells as cellular delivery vehicles for prodrug gene therapy of glioblastoma. *Hum Gene Ther* 2011; 22: 1365-1377.
- [37] Kim MH, Billiar TR, Seol DW. The secretable form of trimeric TRAIL, a potent inducer of apoptosis. *Biochem Biophys Res Commun* 2004; 321: 930-935.
- [38] Kim CY, Jeong M, Mushiaki H, Kim BM, Kim WB, Ko JP, Kim MH, Kim M, Kim TH, Robbins PD, Billiar TR, Seol DW. Cancer gene therapy using a novel secretable trimeric TRAIL. *Gene Ther* 2006; 13: 330-338.
- [39] Sasportas LS, Kasmieh R, Wakimoto H, Hingtgen S, van de Water JA, Mohapatra G, Figueiredo JL, Martuza RL, Weissleder R, Shah K. Assessment of therapeutic efficacy and fate of engineered human mesenchymal stem cells for cancer therapy. *Proc Natl Acad Sci U S A* 2009; 106: 4822-4827.
- [40] Choi SA, Hwang SK, Wang KC, Cho BK, Phi JH, Lee JY, Jung HW, Lee DH, Kim SK. Therapeutic efficacy and safety of TRAIL-producing human adipose tissue-derived mesenchymal stem cells against experimental brainstem glioma. *Neuro Oncol* 2011; 13: 61-69.
- [41] Menon LG, Kelly K, Yang HW, Kim SK, Black PM, Carroll RS. Human bone marrow-derived mesenchymal stromal cells expressing S-TRAIL

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- as a cellular delivery vehicle for human glioma therapy. *Stem Cells* 2009; 27: 2320-2330.
- [42] Kucerova L, Altanerova V, Matuskova M, Tyciakova S, Altaner C. Adipose tissue-derived human mesenchymal stem cells mediated pro-drug cancer gene therapy. *Cancer Res* 2007; 67: 6304-6313.
- [43] Altanerova V, Cihova M, Babic M, Rychly B, Ondicova K, Mravec B, Altaner C. Human adipose tissue-derived mesenchymal stem cells expressing yeast cytosinedeaminase: uracil phosphoribosyltransferase inhibit intracerebral rat glioblastoma. *Int J Cancer* 2012; 130: 2455-2463.
- [44] Kim JH, Lee JE, Kim SU, Cho KG. Stereological analysis on migration of human neural stem cells in the brain of rats bearing glioma. *Neurosurgery* 2010; 66: 333-342; discussion 342.
- [45] Jiang H, Lin JJ, Su ZZ, Goldstein NI, Fisher PB. Subtraction hybridization identifies a novel melanoma differentiation associated gene, mda-7, modulated during human melanoma differentiation, growth and progression. *Oncogene* 1995; 11: 2477-2486.
- [46] Huang EY, Madireddi MT, Gopalkrishnan RV, Leszczyniecka M, Su Z, Lebedeva IV, Kang D, Jiang H, Lin JJ, Alexandre D, Chen Y, Vozhilla N, Mei MX, Christiansen KA, Sivo F, Goldstein NI, Mhashilkar AB, Chada S, Huberman E, Pestka S, Fisher PB. Genomic structure, chromosomal localization and expression profile of a novel melanoma differentiation associated (mda-7) gene with cancer specific growth suppressing and apoptosis inducing properties. *Oncogene* 2001; 20: 7051-7063.
- [47] Caudell EG, Mumm JB, Poindexter N, Ekmekcioglu S, Mhashilkar AM, Yang XH, Retter MW, Hill P, Chada S, Grimm EA. The protein product of the tumor suppressor gene, melanoma differentiation-associated gene 7, exhibits immunostimulatory activity and is designated IL-24. *J Immunol* 2002; 168: 6041-6046.
- [48] Gupta P, Su ZZ, Lebedeva IV, Sarkar D, Sauane M, Emdad L, Bachelor MA, Grant S, Curiel DT, Dent P, Fisher PB. mda-7/IL-24: multifunctional cancer-specific apoptosis-inducing cytokine. *Pharmacol Ther* 2006; 111: 596-628.
- [49] Fisher PB, Gopalkrishnan RV, Chada S, Ramesh R, Grimm EA, Rosenfeld MR, Curiel DT, Dent P. mda-7/IL-24, a novel cancer selective apoptosis inducing cytokine gene: from the laboratory into the clinic. *Cancer Biol Ther* 2003; 2: S23-S37.
- [50] Cunningham CC, Chada S, Merritt JA, Tong A, Senzer N, Zhang Y, Mhashilkar A, Parker K, Vukelja S, Richards D, Hood J, Coffee K, Nemunaitis J. Clinical and local biological effects of an intratumoral injection of mda-7 (IL24; INGN 241) in patients with advanced carcinoma: a phase I study. *Mol Ther* 2005; 11: 149-159.
- [51] Fisher PB. Is mda-7/IL-24 a "magic bullet" for cancer? *Cancer Res* 2005; 65: 10128-10138.
- [52] Su Z, Lebedeva IV, Gopalkrishnan RV, Goldstein NI, Stein CA, Reed JC, Dent P, Fisher PB. A combinatorial approach for selectively inducing programmed cell death in human pancreatic cancer cells. *Proc Natl Acad Sci U S A* 2001; 98: 10332-10337.
- [53] Su ZZ, Madireddi MT, Lin JJ, Young CS, Kitada S, Reed JC, Goldstein NI, Fisher PB. The cancer growth suppressor gene mda-7 selectively induces apoptosis in human breast cancer cells and inhibits tumor growth in nude mice. *Proc Natl Acad Sci U S A* 1998; 95: 14400-14405.
- [54] Emdad L, Lebedeva IV, Su ZZ, Gupta P, Sauane M, Dash R, Grant S, Dent P, Curiel DT, Sarkar D, Fisher PB. Historical perspective and recent insights into our understanding of the molecular and biochemical basis of the antitumor properties of mda-7/IL-24. *Cancer Biol Ther* 2009; 8: 391-400.
- [55] Gupta P, Walter MR, Su ZZ, Lebedeva IV, Emdad L, Randolph A, Valerie K, Sarkar D, Fisher PB. BiP/GRP78 is an intracellular target for MDA-7/IL-24 induction of cancer-specific apoptosis. *Cancer Res* 2006; 66: 8182-8191.
- [56] Hamed HA, Yacoub A, Park MA, Eulitt PJ, Dash R, Sarkar D, Dmitriev IP, Lesniak MS, Shah K, Grant S, Curiel DT, Fisher PB, Dent P. Inhibition of multiple protective signaling pathways and Ad.5/3 delivery enhances mda-7/IL-24 therapy of malignant glioma. *Mol Ther* 2010; 18: 1130-1142.
- [57] Debinski W, Pastan I. An immunotoxin with increased activity and homogeneity produced by reducing the number of lysine residues in recombinant *Pseudomonas* exotoxin. *Bioconjug Chem* 1994; 5: 40-46.
- [58] Okada H, Low KL, Kohanbash G, McDonald HA, Hamilton RL, Pollack IF. Expression of glioma-associated antigens in pediatric brain stem and non-brain stem gliomas. *J Neurooncol* 2008; 88: 245-250.
- [59] Debinski W, Slagle B, Gibo DM, Powers SK, Gillespie GY. Expression of a restrictive receptor for interleukin 13 is associated with glial transformation. *J Neurooncol* 2000; 48: 103-111.
- [60] Candolfi M, Xiong W, Yagiz K, Liu C, Muhammad AK, Puntel M, Foulad D, Zadmehr A, Ahlzadeh GE, Kroeger KM, Tesarfreund M, Lee S, Debinski W, Sareen D, Svendsen CN, Rodriguez R, Lowenstein PR, Castro MG. Gene therapy-mediated delivery of targeted cytotoxins for glioma therapeutics. *Proc Natl Acad Sci U S A* 2010; 107: 20021-20026.

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- [61] Surawska H, Ma PC, Salgia R. The role of ephrins and Eph receptors in cancer. *Cytokine Growth Factor Rev* 2004; 15: 419-433.
- [62] Sun XL, Xu ZM, Ke YQ, Hu CC, Wang SY, Ling GQ, Yan ZJ, Liu YJ, Song ZH, Jiang XD, Xu RX. Molecular targeting of malignant glioma cells with an EphA2-specific immunotoxin delivered by human bone marrow-derived mesenchymal stem cells. *Cancer Lett* 2011; 312: 168-177.
- [63] Yang I, Han SJ, Kaur G, Crane C, Parsa AT. The role of microglia in central nervous system immunity and glioma immunology. *J Clin Neurosci* 2010; 17: 6-10.
- [64] Yang I, Han SJ, Kaur G, Crane C, Parsa AT. The role of microglia in central nervous system immunity and glioma immunology. *J Clin Neurosci* 2010; 17: 6-10.
- [65] Dunn GP, Fecci PE, Curry WT. Cancer immunotherapy in malignant glioma. *Neurosurgery* 2012; 71: 201-222; discussion 222-3.
- [66] Del VM, Bajetta E, Canova S, Lotze MT, Wesa A, Parmiani G, Anichini A. Interleukin-12: biological properties and clinical application. *Clin Cancer Res* 2007; 13: 4677-4685.
- [67] Markert JM, Cody JJ, Parker JN, Coleman JM, Price KH, Kern ER, Quenelle DC, Lakeman AD, Schoeb TR, Palmer CA, Cartner SC, Gillespie GY, Whitley RJ. Preclinical evaluation of a genetically engineered herpes simplex virus expressing interleukin-12. *J Virol* 2012; 86: 5304-5313.
- [68] Ryu CH, Park SH, Park SA, Kim SM, Lim JY, Jeong CH, Yoon WS, Oh WI, Sung YC, Jeun SS. Gene therapy of intracranial glioma using interleukin 12-secreting human umbilical cord blood-derived mesenchymal stem cells. *Hum Gene Ther* 2011; 22: 733-743.
- [69] Breitbach CJ, Thorne SH, Bell JC, Kirn DH. Targeted and armed oncolytic poxviruses for cancer: the lead example of JX-594. *Curr Pharm Biotechnol* 2012; 13: 1768-1772.
- [70] Lun X, Chan J, Zhou H, Sun B, Kelly JJ, Stechishin OO, Bell JC, Parato K, Hu K, Vaillant D, Wang J, Liu TC, Breitbach C, Kirn D, Senger DL, Forsyth PA. Efficacy and safety/toxicity study of recombinant vaccinia virus JX-594 in two immunocompetent animal models of glioma. *Mol Ther* 2010; 18: 1927-1936.
- [71] Drexler HG, Quentmeier H. FLT3: receptor and ligand. *Growth Factors* 2004; 22: 71-73.
- [72] Ali S, Curtin JF, Zirger JM, Xiong W, King GD, Barcia C, Liu C, Puntel M, Goverdhan S, Lowenstein PR, Castro MG. Inflammatory and anti-glioma effects of an adenovirus expressing human soluble Fms-like tyrosine kinase 3 ligand (hsFlt3L): treatment with hsFlt3L inhibits intracranial glioma progression. *Mol Ther* 2004; 10: 1071-1084.
- [73] Barnard Z, Wakimoto H, Zaupa C, Patel AP, Klehm J, Martuza RL, Rabkin SD, Curry WJ. Expression of FMS-like tyrosine kinase 3 ligand by oncolytic herpes simplex virus type I prolongs survival in mice bearing established syngeneic intracranial malignant glioma. *Neurosurgery* 2012; 71: 741-748; discussion 748.
- [74] Rowntree RK, McNeish JD. Induced pluripotent stem cells: opportunities as research and development tools in 21st century drug discovery. *Regen Med* 2010; 5: 557-568.
- [75] Slettenaar VI, Wilson JL. The chemokine network: a target in cancer biology? *Adv Drug Deliv Rev* 2006; 58: 962-974.
- [76] Ebert LM, Schaeferli P, Moser B. Chemokine-mediated control of T cell traffic in lymphoid and peripheral tissues. *Mol Immunol* 2005; 42: 799-809.
- [77] Auffinger B, Morshed R, Tobias A, Cheng Y, Ahmed AU, Lesniak MS. Drug-loaded nanoparticle systems and adult stem cells: a potential marriage for the treatment of malignant glioma? *Oncotarget* 2013; 4: 378-396.
- [78] Gursel DB, Berry N, Boockvar JA. Therapeutic stem cells encapsulated in a synthetic extracellular matrix selectively kill tumor cells, delay tumor growth, and increase survival in a mouse resection model of malignant glioma. *Neurosurgery* 2012; 70: N17-N19.
- [79] Mattis VB, Svendsen CN. Induced pluripotent stem cells: a new revolution for clinical neurology? *Lancet Neurol* 2011; 10: 383-394.
- [80] Lee EX, Lam DH, Wu C, Yang J, Tham CK, Ng WH, Wang S. Glioma gene therapy using induced pluripotent stem cell derived neural stem cells. *Mol Pharm* 2011; 8: 1515-1524.
- [81] Yamanaka S. A fresh look at iPS cells. *Cell* 2009; 137: 13-17.